

Bioinformatics Analysis in R

Advanced Gene Expression: Analysis of Cancer Genome Atlas

Ivan G. Costa, Martin Grasshoff

Institute for Computational Genomics
RWTH University Hospital
www.costalab.org

Summary

1. Obtain data from cancer patients from TCGA
2. Pre-process and analysis of RNA-seq data
3. Use machine learning to build a classifier for personalised medicine
4. Use interesting markers for survival analysis

The Cancer Genome Atlas

- TCGA is a NCI (US) funded project to generate cohorts of cancers:
 - Currently 33 cancers with 80-780 patients
- Comprehensive data from tissues:
 - Histology, clinical, gene expression profiling, copy number variation, DNA methylation using arrays or sequencing
- Data is publicly available upon generation and deposited in a portal (portal.gdc.cancer.gov)

The Cancer Genome Atlas - Portal

Harmonized Cancer Datasets

Genomic Data Commons Data Portal

Get Started by Exploring:

- Projects
- Exploration
- Analysis
- Repository

Q e.g. BRAF, Breast, TCGA-BLCA, TCGA-A5-A0G2

Data Portal Summary

Data Release 13.0 - September 27, 2018

PROJECTS

43

PRIMARY SITES

69

CASES

33,096

FILES

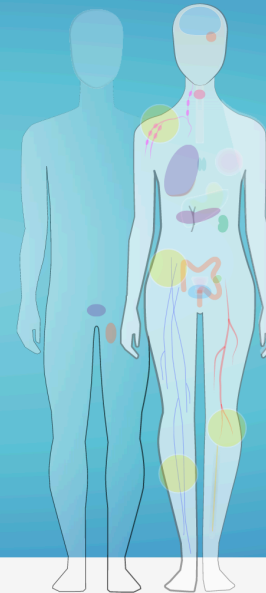
358,092

GENES

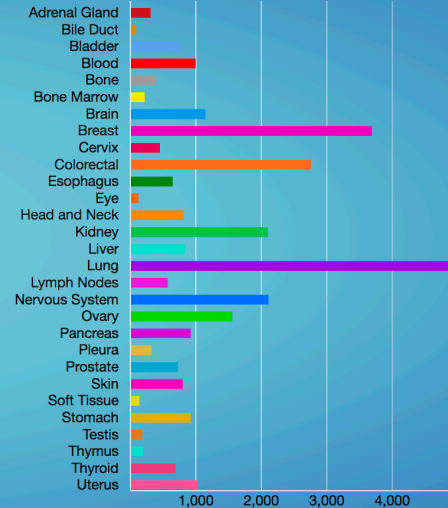
22,147

MUTATIONS

3,142,246

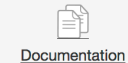
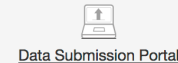
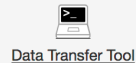


Cases by Major Primary Site



GDC Applications

The GDC Data Portal is a robust data-driven platform that allows cancer researchers and bioinformaticians to search and download cancer data for analysis. The GDC applications include:



The Cancer Genome Atlas - Portal

NIH NATIONAL CANCER INSTITUTE GDC Data Portal

Home Projects Exploration Analysis Repository

Quick Search Manage Sets Login Cart 0 GDC Apps

Harmonized Cancer Datasets

Genomic Data Commons Data Portal

Get Started by Exploring:

Projects Exploration Analysis Repository

Q e.g. BRAF, Breast, TCGA-BLCA, TCGA-A5-A0G2

Data Portal Summary

Data Release 13.0 - September 27, 2018

PROJECTS 43	PRIMARY SITES 69	CASES 33,096
FILES 358,092	GENES 22,147	MUTATIONS 3,142,246

Cases by Major Primary Site

Primary Site	Cases
Adrenal Gland	~100
Bile Duct	~100
Bladder	~100
Blood	~100
Bone	~100
Bone Marrow	~100
Brain	~100
Breast	~3,500
Cervix	~100
Colorectal	~2,500
Esophagus	~100
Eye	~100
Head and Neck	~100
Kidney	~100
Liver	~100
Lung	~4,500
Lymph Nodes	~100
Nervous System	~2,000
Ovary	~100
Pancreas	~100
Pleura	~100
Prostate	~100
Skin	~100
Soft Tissue	~100
Stomach	~100
Testis	~100
Thymus	~100
Thyroid	~100
Uterus	~100

GDC Applications

The GDC Data Portal is a robust data-driven platform that allows cancer researchers and bioinformaticians to search and download cancer data for analysis. The GDC applications include:

- Data Portal
- Website
- Data Transfer Tool
- API
- Data Submission Portal
- Documentation
- Legacy Archive

Check a gene or cancer type!
I will try liver

LIHC - Liver Hepatocellular Carcinoma

[Explore Project Data](#)
[Biospecimen](#)
[Clinical](#)
[Manifest](#)

Summary

Project ID	TCGA-LIHC
Project Name	Liver Hepatocellular Carcinoma
Disease Type	Adenomas and Adenocarcinomas
Primary Site	Liver and intrahepatic bile ducts
Program	TCGA

CASES

[377](#)


FILES

[10,814](#)


ANNOTATIONS

[28](#)


Cases and File Counts by Data Category

Data Category	Cases (n=377)	Files (n=10,814)
Raw Sequencing Data	377	1,637
Transcriptome Profiling	376	2,122
Simple Nucleotide Variation	375	3,032
Copy Number Variation	376	1,536
DNA Methylation	377	430
Clinical	377	423
Biospecimen	377	1,634

Cases and File Counts by Experimental Strategy

Experimental Strategy	Cases (n=377)	Files (n=10,814)
Diagnostic Slide	365	379
Tissue Slide	377	491
WXS	376	3,820
RNA-Seq	371	1,696
miRNA-Seq	373	1,275
Genotyping Array	376	1,536
Methylation Array	377	430

LIHC - Liver Hepatocellular Carcinoma

Explore Project Data

Biospecimen

Clinical

Manifest

Summary

Project ID	TCGA-LIHC
Project Name	Liver Hepatocellular Carcinoma
Disease Type	Adenomas and Adenocarcinomas
Primary Site	Liver and intrahepatic bile ducts
Program	TCGA

CASES
[377](#)



FILES
[10,814](#)



ANNOTATIONS
[28](#)



Cases and File Counts by Data Category

Data Category	Cases (n=377)	Files (n=10,814)
Raw Sequencing Data	377	1,637
Transcriptome Profiling	376	2,122
Simple Nucleotide Variation	375	3,032
Copy Number Variation	376	1,536
DNA Methylation	377	430
Clinical	377	423
Biospecimen	377	1,634

Cases and File Counts by Experimental Strategy

Experimental Strategy	Cases (n=377)	Files (n=10,814)
Diagnostic Slide	365	379
Tissue Slide	377	491
WXS	376	3,820
RNA-Seq	371	1,696
miRNA-Seq	373	1,275
Genotyping Array	376	1,536
Methylation Array	377	430

Gene expression data!

LIHC - Liver Hepatocellular Carcinoma

Files Cases

[Add a File Filter](#)

File

Q e.g. 142682.bam, 4f6e2e7a-b...

Data Category

- Simple Nucleotide Variation 3,032
- Transcriptome Profiling 2,122
- Raw Sequencing Data 1,637
- Biospecimen 1,634
- Copy Number Variation 1,536

2 More...

Data Type

- Gene Expression Quantification 1,272
- Isoform Expression Quantification 425
- miRNA Expression Quantification 425

Experimental Strategy

- RNA-Seq 1,272
- miRNA-Seq 860

Workflow Type

- BCGSC miRNA Profiling 860
- HTSeq - Counts 424
- HTSeq - FPKM 424
- HTSeq - FPKM-UQ 424

Data Format

- TXT 2,122

Platform

No data for this field

Access


- open 2,122

Clear
Project Id
IS
TCGA-LIHC
AND
Data Category
IS
Transcriptome Profiling


Add All Files to Cart
Manifest
View 376 Cases in Exploration
View Images

Files (2,122)
Cases (376)


Primary Site



Project



Data Category



[Show More](#)

Showing 1 - 20 of 2,122 files

	Access	File Name	Cases	Project
	open	7085ee3a-b604-4a12-a877-63eef2d905e8.htseq.counts.gz	1	TCGA-LIHC
	open	acf3d05a-0ca4-4fee-8f07-44b93017b5fd.mirbase21.isoforms.quantification.txt	1	TCGA-LIHC
	open	13240f8b-ae36-4f5f-8e95-2c9d0c83e58c.FPKM-UQ.txt.gz	1	TCGA-LIHC
	open	77e29a20-68d3-4881-a3ac-a564359bcc05.FPKM-UQ.txt.gz	1	TCGA-LIHC
	open	103b1320-8c4e-44ea-9449-fdcb6b405f94.htseq.counts.gz	1	TCGA-LIHC
	open	466776cb-6906-4da2-b788-a05a154decf3.mirbase21.mirnas.quantification.txt	1	TCGA-LIHC
	open	e4c90512-0e06-4517-95fe-c10b999f5f81.mirbase21.mirnas.quantification.txt	1	TCGA-LIHC
	open	5f94c33f-588b-4b6a-9c13-4505b0f94403.htseq.counts.gz	1	TCGA-LIHC
	open	6ce06871-a6a4-4a4a-bd08-0c448914dfcf.FPKM.txt.gz	1	TCGA-LIHC
	open	a762a98f-9041-47e2-8561-46fae396f12.htseq.counts.gz	1	TCGA-LIHC
	open	61ec8919-8b12-43d7-b127-8b68a66bd033.mirbase21.mirnas.quantification.txt	1	TCGA-LIHC
	open	f3e152ef-5048-4157-a195-d13ed8851170.htseq.counts.gz	1	TCGA-LIHC
	open	ca28f37f-d686-41f9-90fb-9da55fec40cb.mirbase21.isoforms.quantification.txt	1	TCGA-LIHC
	open	13240f8b-ae36-4f5f-8e95-2c9d0c83e58c.FPKM.txt.gz	1	TCGA-LIHC
	open	e035a46e-6114-4a64-b5ae-9e6209223493.FPKM.txt.gz	1	TCGA-LIHC
	open	a96f2f6c-38e0-453c-961d-aa83b92652da.mirbase21.mirnas.quantification.txt	1	TCGA-LIHC
	open	a0c56eec-568a-46b0-88db-f14d64a3942b.FPKM.txt.gz	1	TCGA-LIHC
	open	9c644f65-0ebb-4862-98a9-308b81c8fb26.mirbase21.mirnas.quantification.txt	1	TCGA-LIHC
	open	ad114591-0409-4bc5-8f0b-dbb44a5ad0eb.mirbase21.isoforms.quantification.txt	1	TCGA-LIHC
	open	3edd413e-831d-442a-be8d-70b2f49e9d67.FPKM.txt.gz	1	TCGA-LIHC

Show 20 entries

LIHC - Liver Hepatocellular Carcinoma

Files Cases

Clear Project Id IS TCGA-LIHC AND Data Category IS Transcriptome Profiling

Add All Files to Cart Manifest View 376 Cases in Exploration View Images

Files (2,122) Cases (376)

Primary Site Project Data Category

Show More

Showing 1 - 20 of 2,122 files

Access	File Name	Cases	Project
open	7086e931-f8m-412-a1m-67e62-90508.tsc.ccrins.fr	1	TCGA-LIHC
open	acf0c03a-0ca4-4fec-bf07-44b9001b5fd.mirbase21.isoforms.quantification.txt	1	TCGA-LIHC
open	13240f8b-ae36-4f5f-8e95-2c9d0c83e58c.FPKM-UQ.txt.gz	1	TCGA-LIHC
open	77e29a20-68d3-4881-a3ac-a564359bcc05.FPKM-UQ.txt.gz	1	TCGA-LIHC
open	103b1320-8c4e-44ea-9449-fdcb6b405f94.htseq.counts.gz	1	TCGA-LIHC
open	466776cb-6906-4da2-b788-a05a154decf3.mirbase21.mirnas.quantification.txt	1	TCGA-LIHC
open	e4c90512-0e06-4517-95fe-c10b999f5f81.mirbase21.mirnas.quantification.txt	1	TCGA-LIHC
open	5f94c33f-588b-4b6a-9c13-4505b0f94403.htseq.counts.gz	1	TCGA-LIHC
open	6ce06871-a6a4-4a4a-bd08-0c448914dfcf.FPKM.txt.gz	1	TCGA-LIHC
open	a762a98f-9041-47e2-8561-46fae396f12.htseq.counts.gz	1	TCGA-LIHC
open	61ec819-b12-43d7-b127-8b68a661d033.mirbase21.mirnas.quantification.txt	1	TCGA-LIHC
open	f3e15af-5f4c-415c-495d11e1885170.htseq.counts.gz	1	TCGA-LIHC
open	ca28f37f-d686-41f9-90fb-9da55fec40cb.mirbase21.isoforms.quantification.txt	1	TCGA-LIHC
open	13240f8b-ae36-4f5f-8e95-2c9d0c83e58c.FPKM.txt.gz	1	TCGA-LIHC
open	e035a46e-6114-4a64-b5ae-9e6209223493.FPKM.txt.gz	1	TCGA-LIHC
open	a96f2f6c-38e0-453c-961d-aa83b92652da.mirbase21.mirnas.quantification.txt	1	TCGA-LIHC
open	a0c56eec-568a-46b0-88db-f14d64a3942b.FPKM.txt.gz	1	TCGA-LIHC
open	9c644f65-0ebb-4862-98a9-308b81c8fb26.mirbase21.mirnas.quantification.txt	1	TCGA-LIHC
open	ad114591-0409-4bc5-8f0b-dbb445ad0eb.mirbase21.isoforms.quantification.txt	1	TCGA-LIHC
open	3edd413e-831d-442a-be8d-70b2f49e9d67.FPKM.txt.gz	1	TCGA-LIHC

Show 20 entries

Distinct ways to represent transcripts

Distinct ways to count gene expression.

Data Category

- Simple Nucleotide Variation (3,032)
- Transcriptome Profiling (2,122)**
- Raw Sequencing Data (1,637)
- Biospecimen (1,634)
- Copy Number Variation (1,536)

Data Type

- Gene Expression Quantification (1,272)
- Isoform Expression Quantification (425)
- miRNA Expression Quantification (425)

Experimental Strategy

- RNA-Seq (1,272)
- miRNA-Seq (860)

Workflow Type

- BCGSC miRNA Profiling (860)
- HTSeq - Counts (424)
- HTSeq - FPKM (424)
- HTSeq - FPKM-UQ (424)

Data Format

- TEXT (2,122)

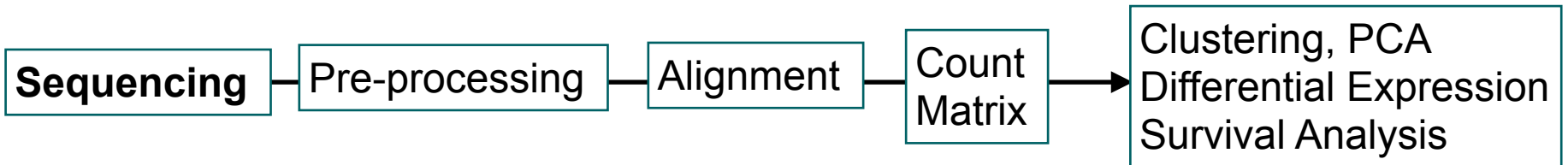
Platform

No data for this field

Access

- open (2,122)

Bioinformatics Pipeline / RNA-seq



Bioinformatics Pipeline / RNA-seq



Sequencing

Pre-processing

Alignment

Count
Matrix

Clustering, PCA
Differential Expression
Survival Analysis

Practical part not covered!

Bioinformatics Pipeline / RNA-seq



Sequencing

Pre-processing

Alignment

Count
Matrix

Clustering, PCA
Differential Expression
Survival Analysis

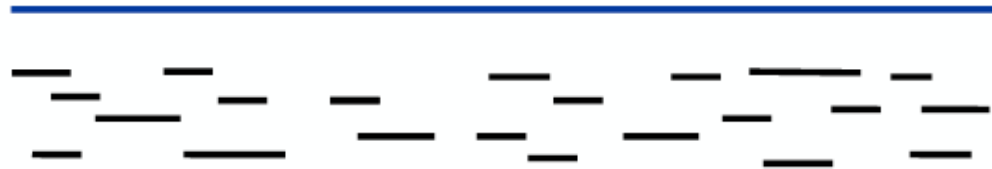
Next Generation Sequencing

- ▶ NGS take advantage of **parallelization**
 - ▶ reads millions/billions of reads per run
 - ▶ short reads (50-100 bps)
 - ▶ error rates (0.1-1%)



Read Types

Fragment DNA:

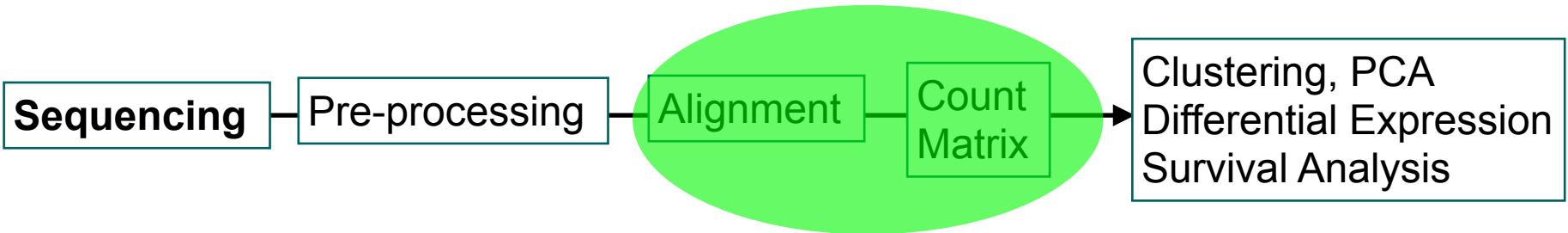


Single end



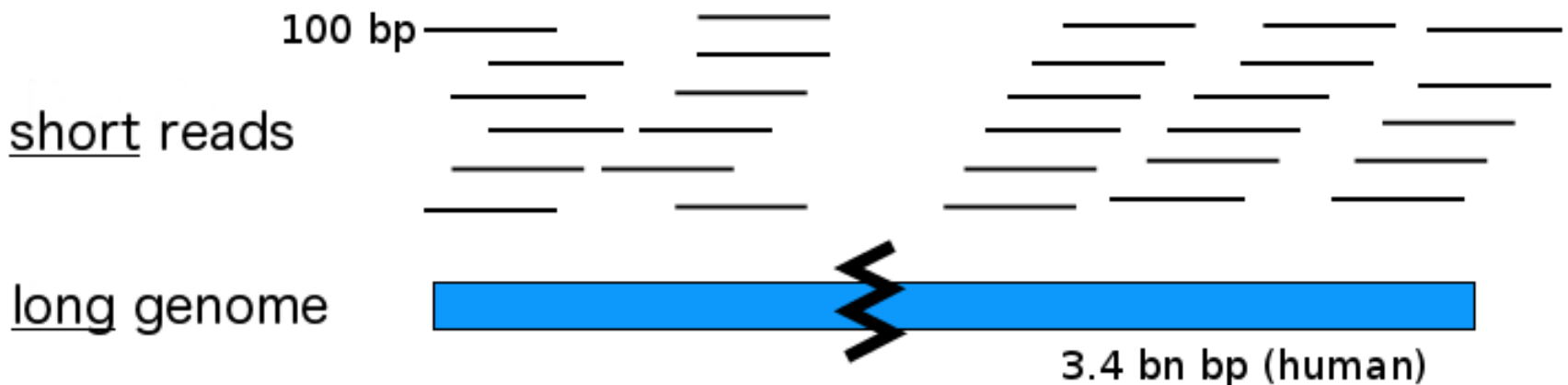
Paired end
Ins: 200-800 bp

Bioinformatics Pipeline / RNA-seq

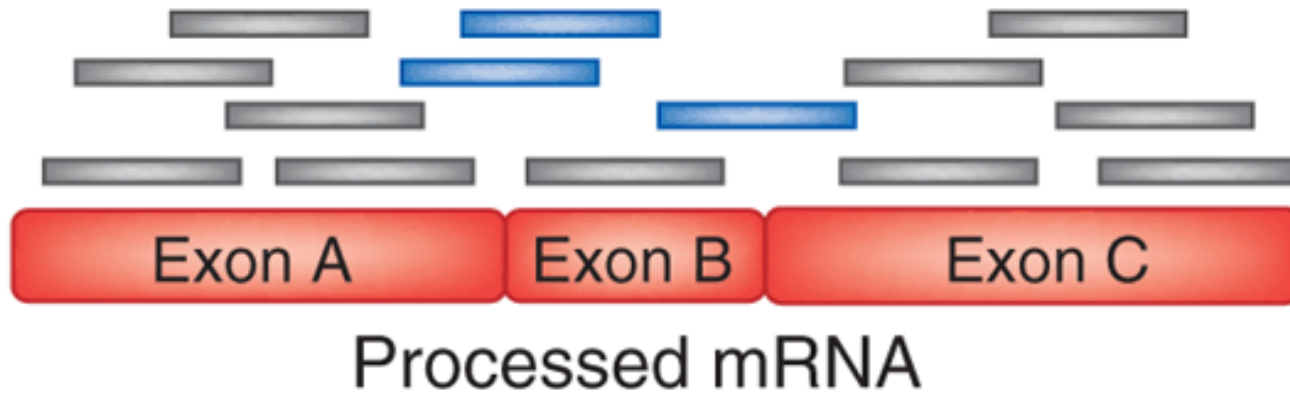


Alignment

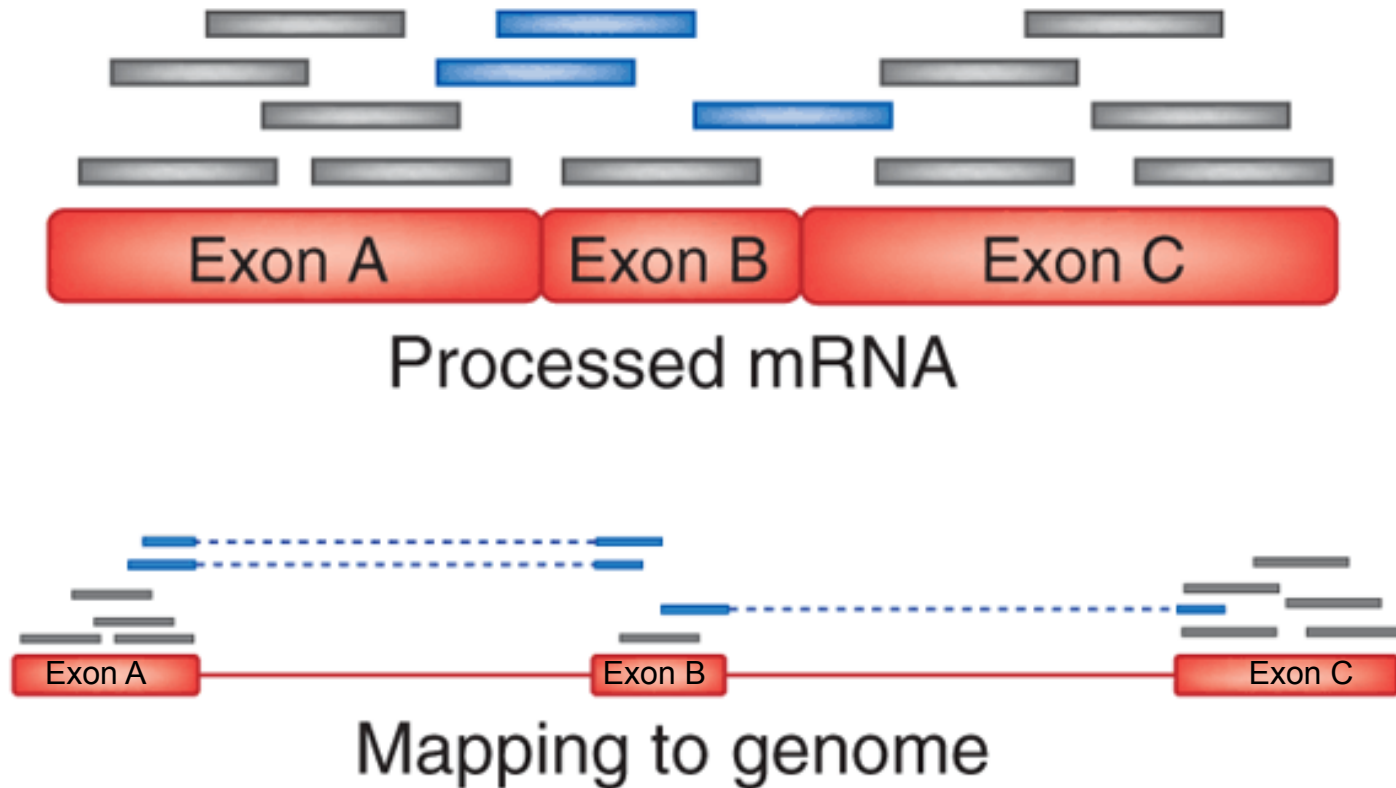
- a large reference sequence is given (genome)
 - up to billions of base pairs
- short reads (<200bps)
- find most probable position of the read in the genome (by inexact string matching)



Alignment - Split Read Mapping (RNA-Seq)



Alignment - Split Read Mapping (RNA-Seq)



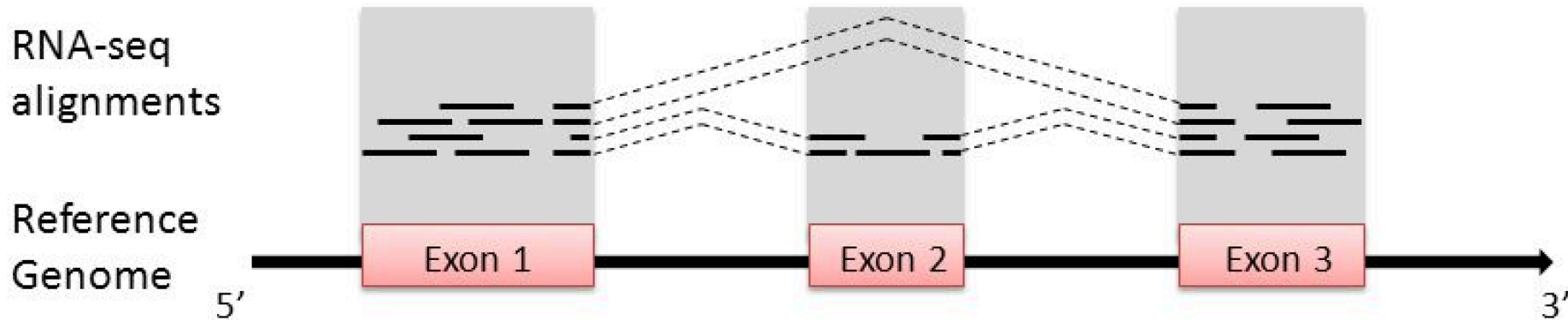
- reads are split between exons when mapped to genome
- aligners use transcript information or try to find splice events (STAR & TOPHAT)

Reference based aligners - Overview

	<i>Time</i>	<i>Precision</i>	<i>Pairs</i>	<i>GAPS</i>	<i>Phred</i>	<i>Memory</i>	<i>Application (Comments)</i>
BOWTIE	+		+	-	-	5GB	General <i>(max. 3 missmatches)</i>
BWA	+		+	+	+	8GB	General <i>(max of 200bps reads)</i>
NOVOALIGN		+	+	+	+	8GB	General <i>(commercial license)</i>
STAR	+		+	-	+	32GB	RNA-Seq <i>(allow split-maps)</i>
BISMARK	+		+	+	+	10GB	Bisulfite/reduced sequencing

Computers need large memory and a few hours of computation per experiment!

Quantification (Count Matrix)



Simple Counting Approaches

Gene Level - 17 reads

Exon level - exon 1 (8 reads), exon 2 (3 reads), exon 3 (6 reads)

Transcript Level - Exons 1,2 & 3 (10 reads) and exon 1 & 3 (7 reads) *

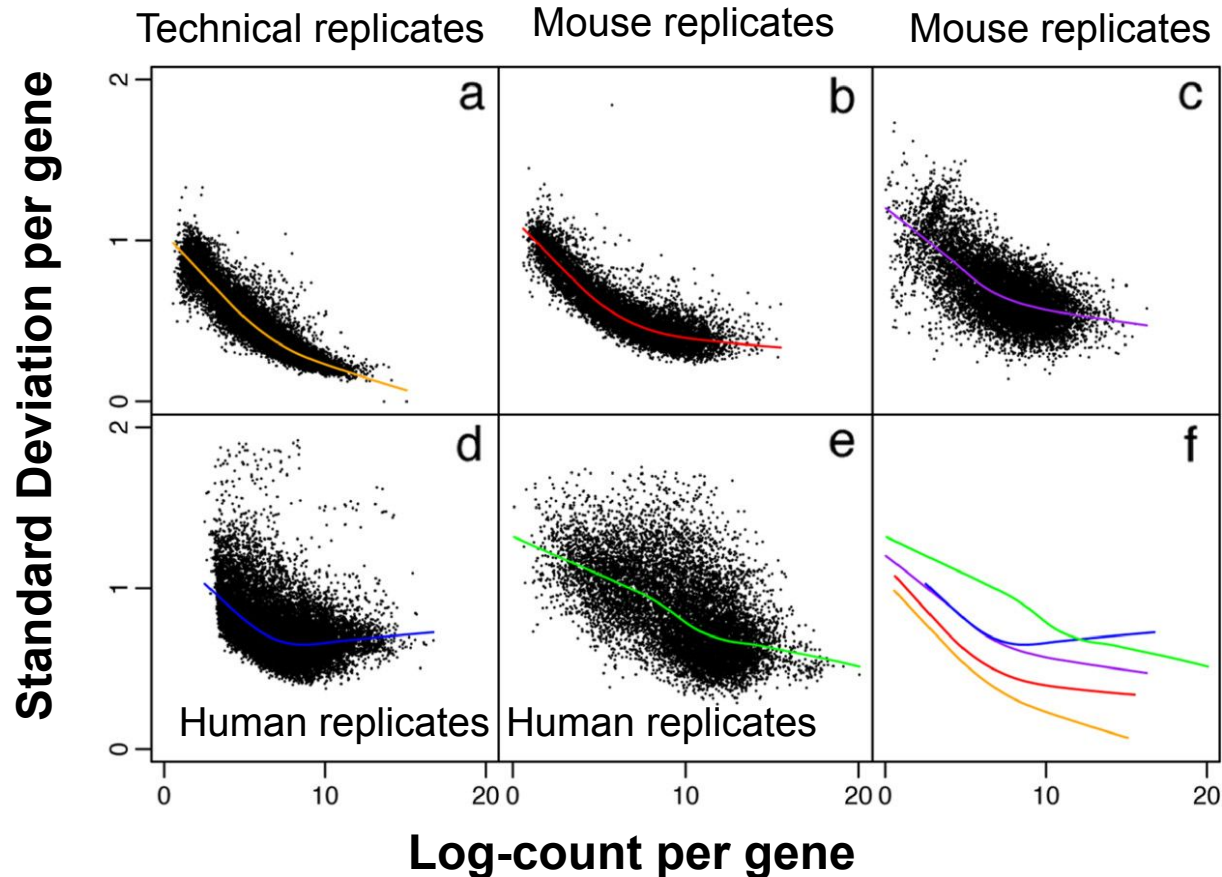
* complex computational methods required (RSe, or TopHAT needed for this)

Fragments per Kilobase (FPKM)

- normalize counts by read size (kb) and RNA-seq library size (mb)

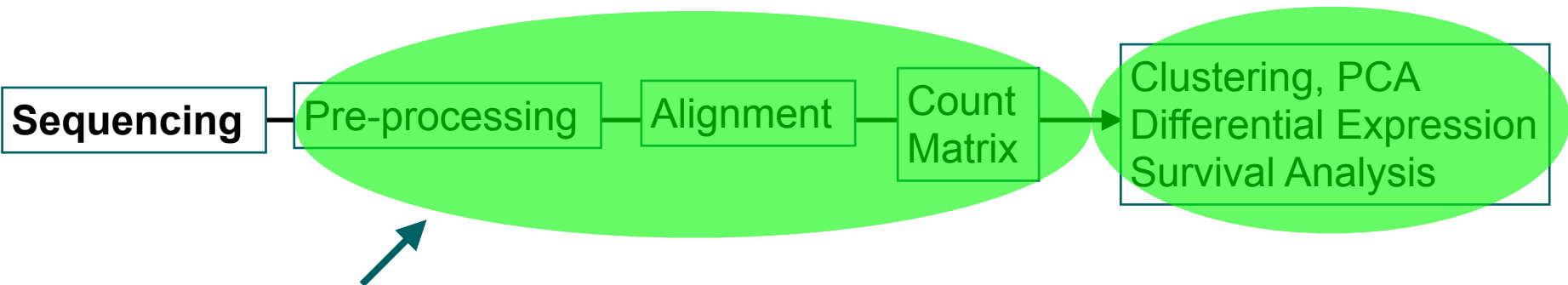
RNA-seq and Differential Analysis

Arrays and RNA-seq have distinct distributions



VOOM analysis is necessary to make variance similar to arrays.

Bioinformatics Pipeline / RNA-seq



Provided by TGCA or your Core Facility!

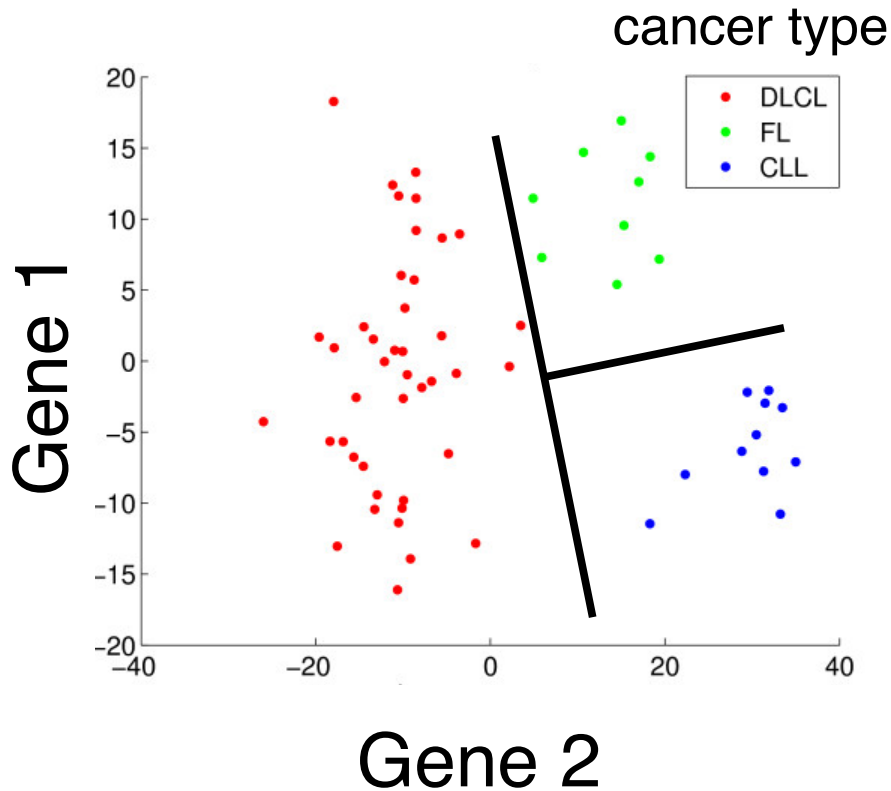
Personalized Medicine

Diagnosis and treatment choices is mostly carried on macromolecular features:

- morphology of tumours (image), symptoms, blood levels

Challenges: use molecular markers (expression or genetics) for diagnosis or treatment selection.

Machine Learning - Classifier



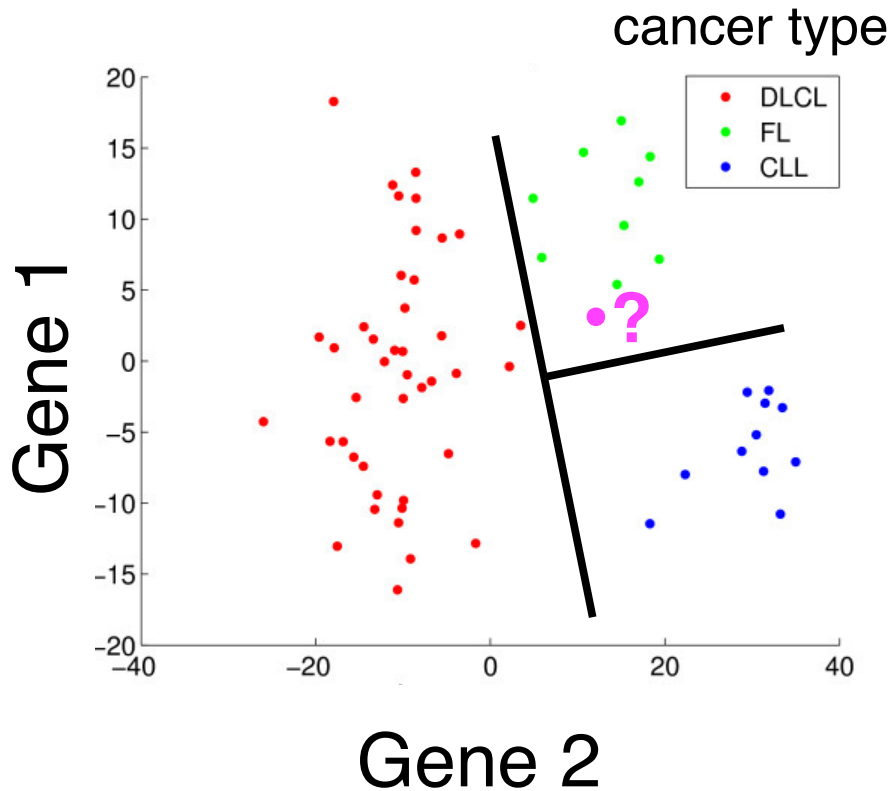
Data

Expression matrix X
(genes vs samples)
classification vector Y
(diagnosis)

Find a function:

$$f(x) \rightarrow y$$

Machine Learning - Classifier



Data

Expression matrix X
(genes vs samples)
classification vector Y
(diagnosis)

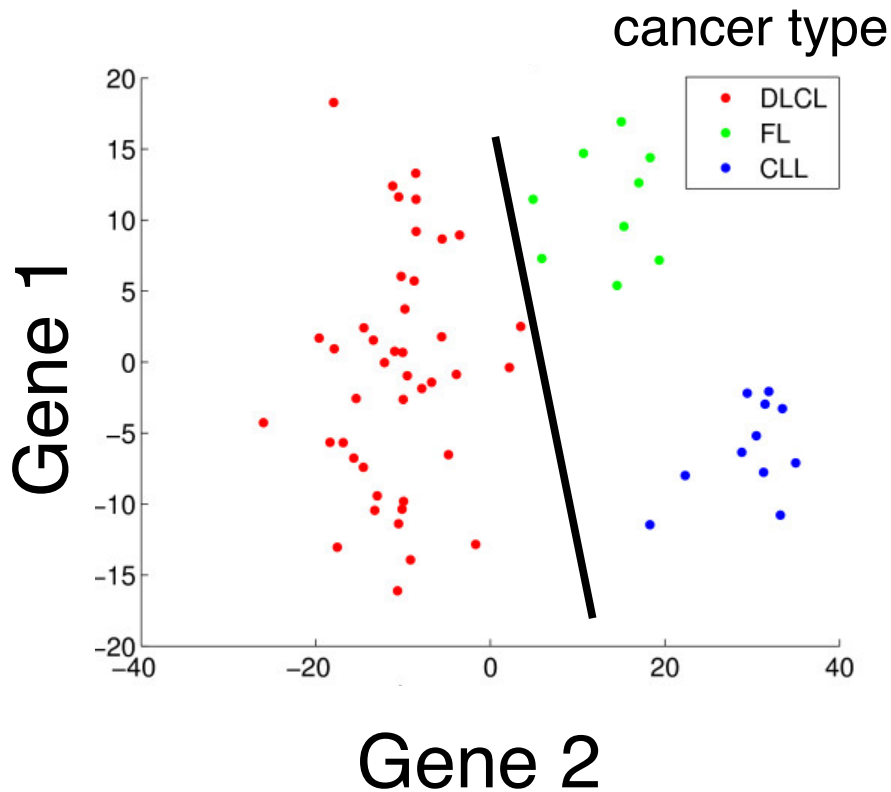
Find a function:

$$f(x) \rightarrow y$$

For new patients X' :

$$f(x') \rightarrow y'$$

Linear Classifier



Linear Function:

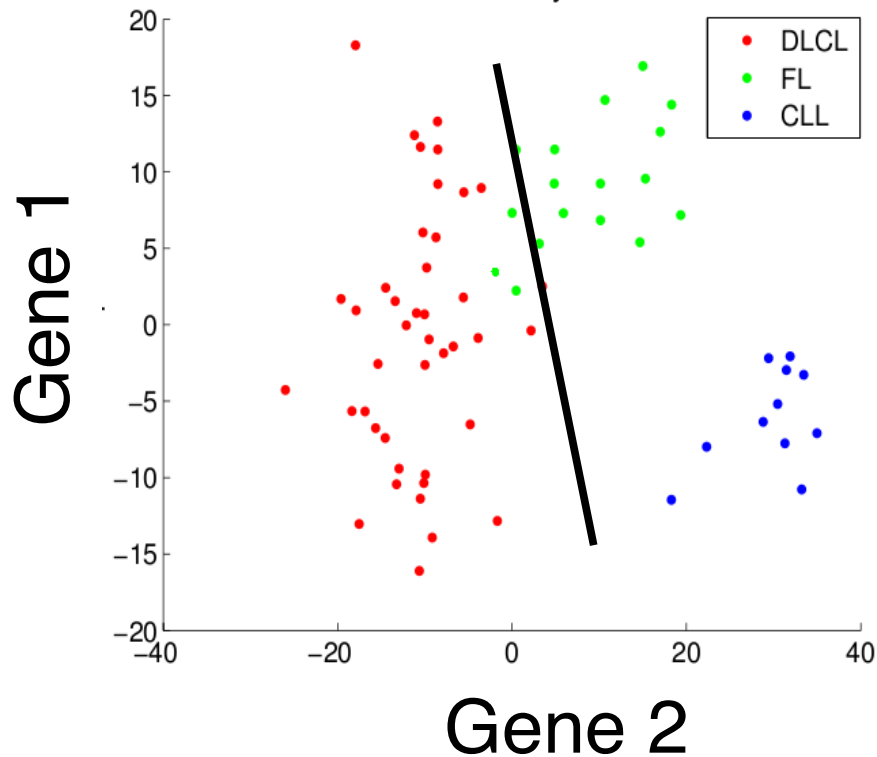
$$f(x, A) = a_0 + a_1x_1 + \dots + a_Lx_L$$

$$f(x, A) > 0 \Rightarrow \text{class A}$$

$$f(x, A) \leq 0 \Rightarrow \text{class B}$$

- Works for 2 classes only
 - Train a function for each cancer type
- Find coefficients A
 - estimated with neural networks or support vector machines

Linear Classifier - Problems



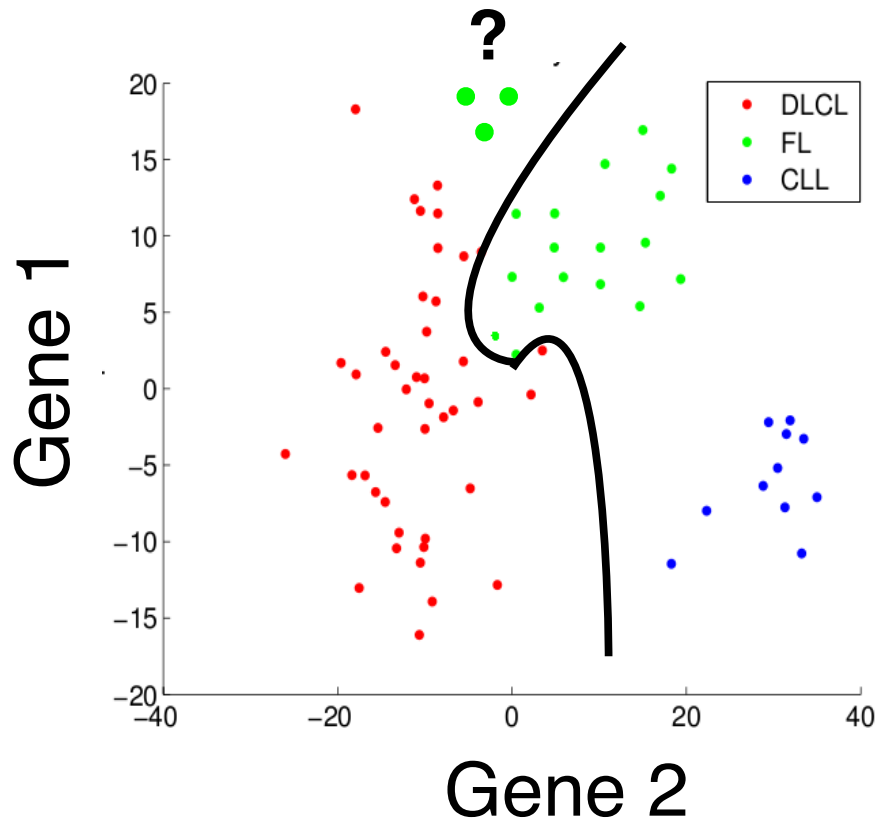
- Most real world problems are not linearly separable!
- There will be always some error!
- Solution: non-linear functions

Nonlinear Classifier - Problems



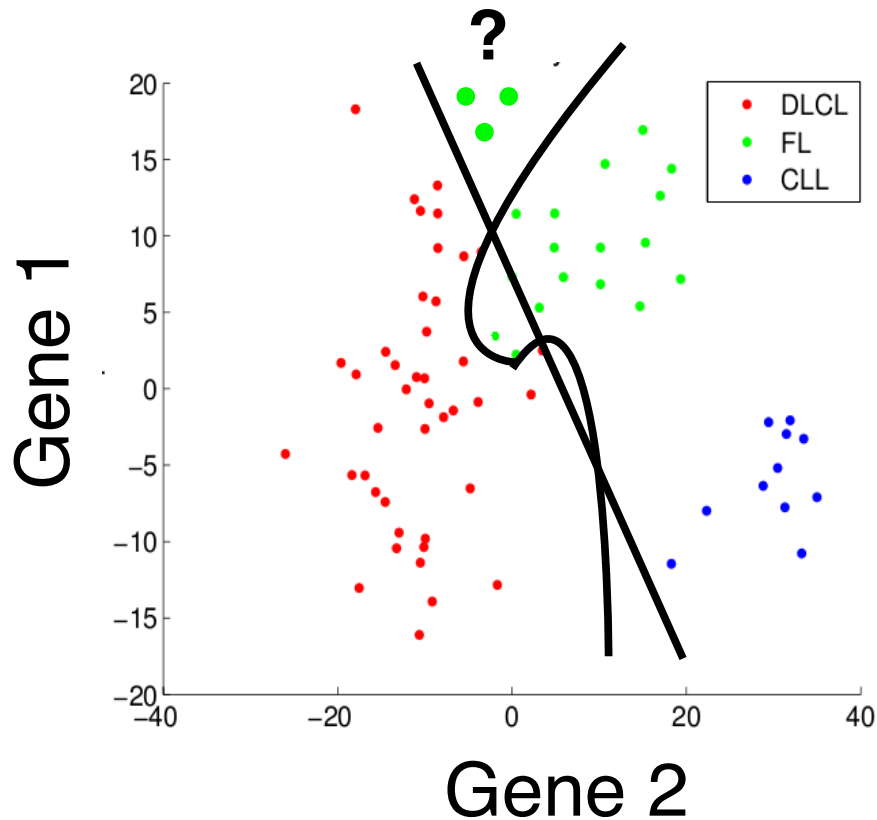
- Polynomial Function
- $f(x, A) = a_0 + a_{11}x^3_1 + \dots + a_{L1}x^3_L$
 $a_{12}x^2_1 + \dots + a_{L2}x^2_L$
 $a_{12}x_1 + \dots + a_{L2}x_L$
- Third order polynomial
- Problem: overfitting

Nonlinear Classifier - Problems



- Polynomial Function
- $f(x, A) = a_0 + a_{11}x^3_1 + \dots + a_{L1}x^3_L$
 $a_{12}x^2_1 + \dots + a_{L2}x^2_L$
 $a_{12}x_1 + \dots + a_{L2}x_L$
- Third order polynomial
- Problem: overfitting

Nonlinear Classifier - Problems



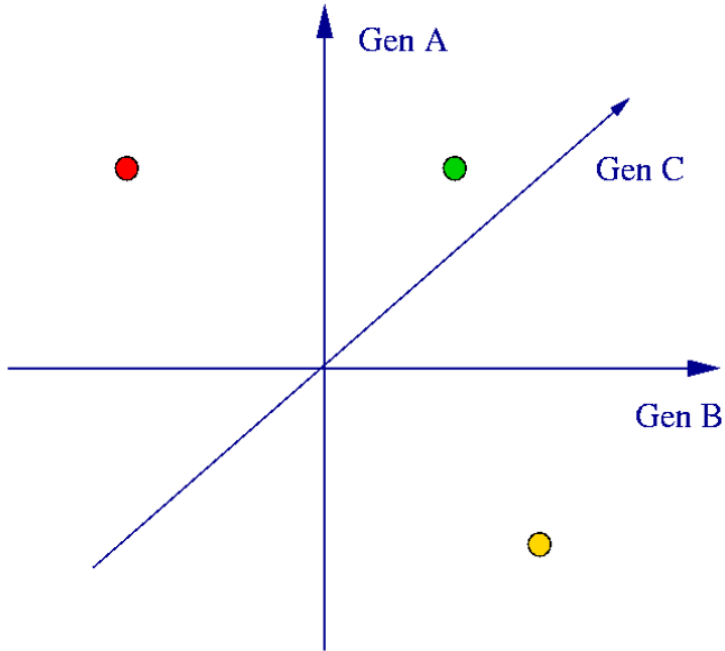
- Polynomial Function
- $f(x, A) = a_0 + a_{11}x^3_1 + \dots + a_{L1}x^3_L$
 $a_{12}x^2_1 + \dots + a_{L2}x^2_L$
 $a_{12}x_1 + \dots + a_{L2}x_L$
- Third order polynomial
- Problem: overfitting

Curse of Dimensionality

Size of a Euclidean space grows with dimension (number of genes)

Dots (patients) are sparsely distributed in space

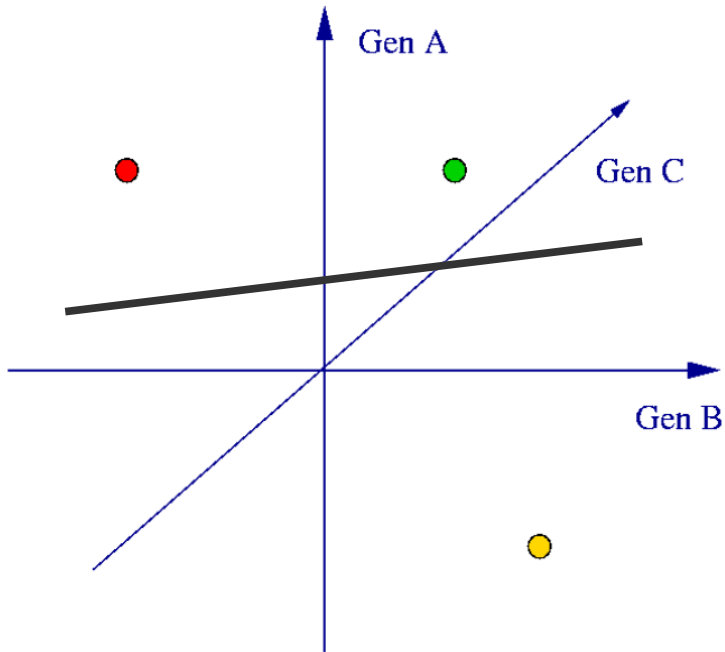
Curse of Dimensionality : Example



Sparse data

- three genes
- 2 patients with known cancer (red vs yellow)
- 1 unknown (green)

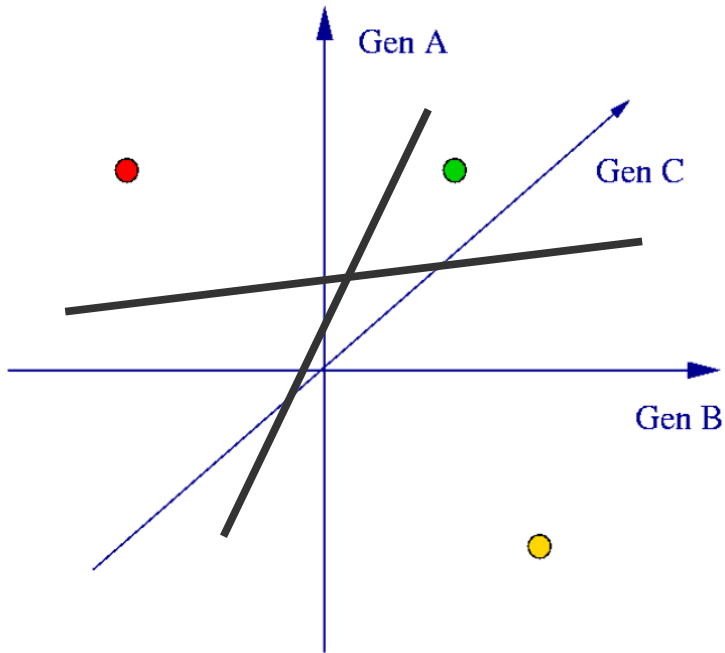
Curse of Dimensionality : Example



- Sparse data
 - three genes
 - 2 patients with known cancer (red vs yellow)
 - 1 unknown (green)

Perfect classifier (on training)

Curse of Dimensionality : Example

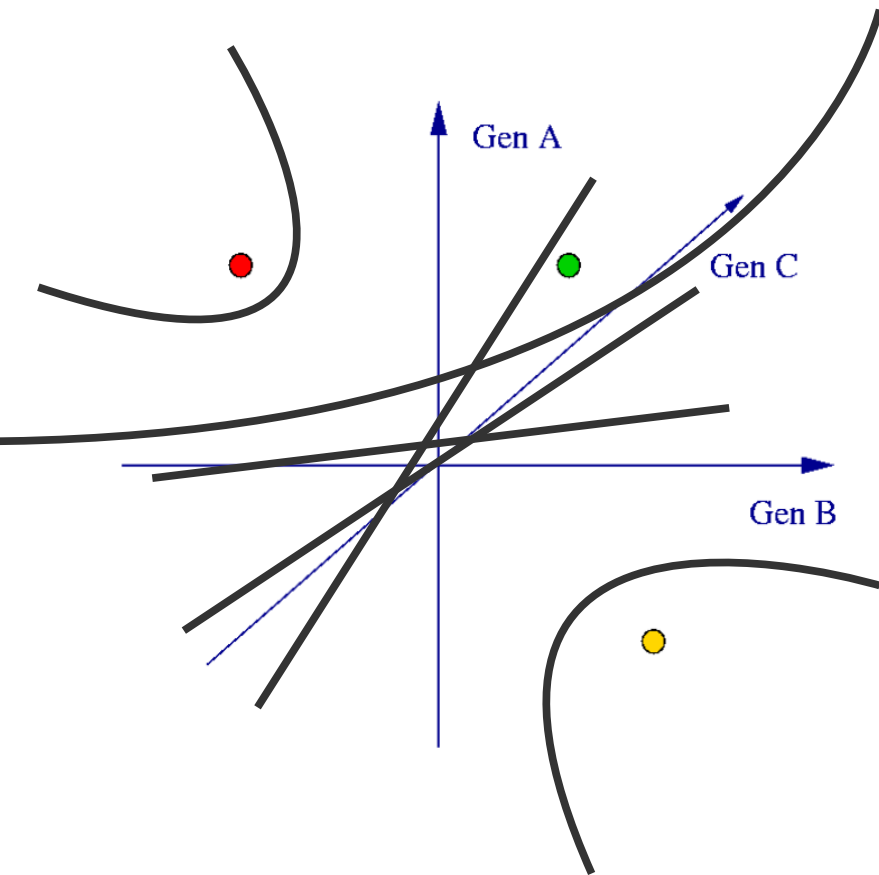


- Sparse data
 - three genes
 - 2 patients with known cancer (red vs yellow)
 - 1 unknown (green)

Both are perfect classifiers
(on training)

Hard to generalise!

Curse of Dimensionality : Example



- There are millions of perfect linear classifiers
- And even more non-linear classifiers!

Dealing with Curse of Dimensionality

- Have a proper training / test evaluation procedure
- Use classifiers which are as simple as possible
- Reduce the dimension of your data (feature selection or PCA)

Classifier Evaluation

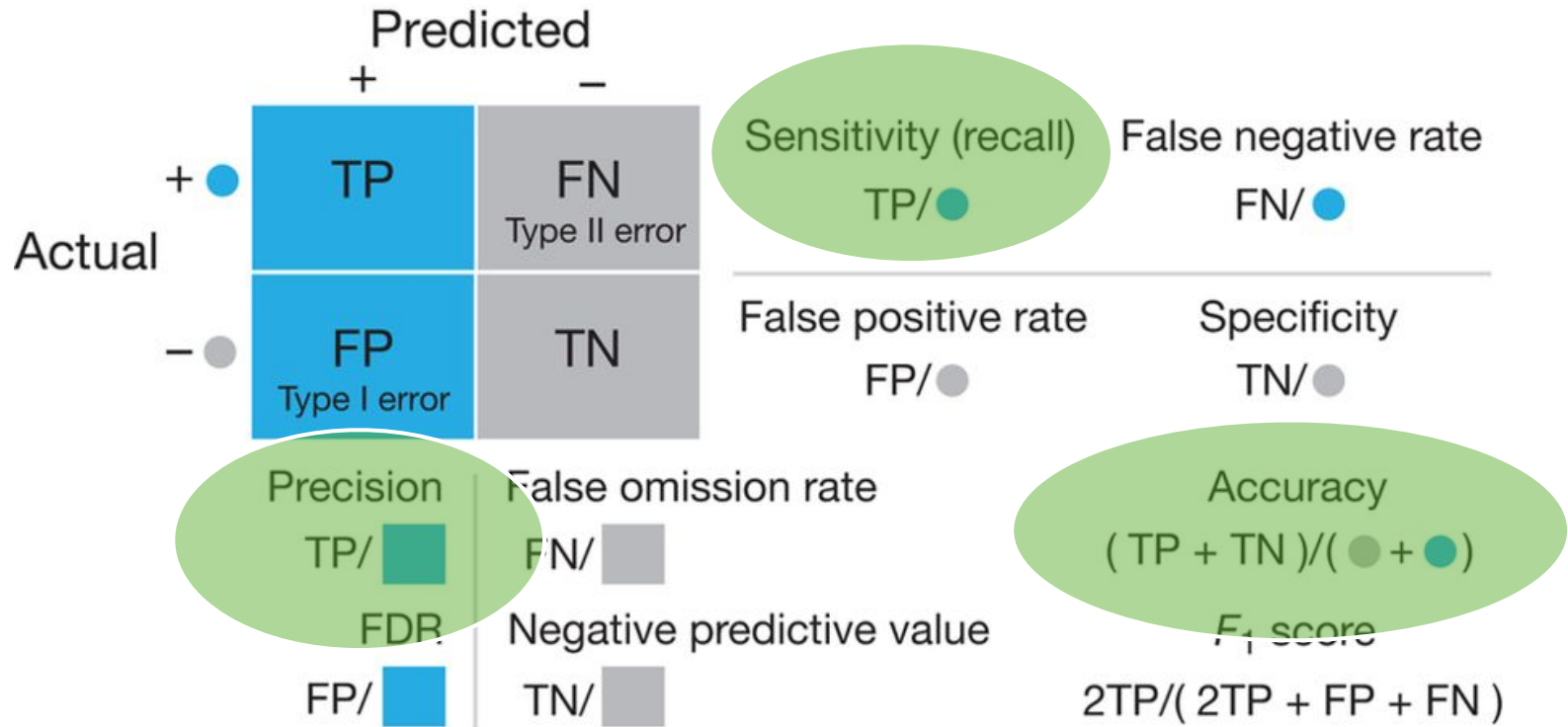
Measures for a two class problem (cancer + vs. non-cancer -)

		Predicted	
		+	-
Actual	+ ●	TP	FN Type II error
	- ●	FP Type I error	TN

Source: Lever et al., Nat. Methods (2016)

Classifier Evaluation

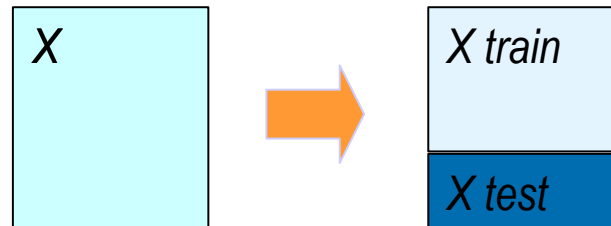
Measures for a two class problem (cancer + vs. non-cancer -)



Source: Lever et al., Nat. Methods (2016)

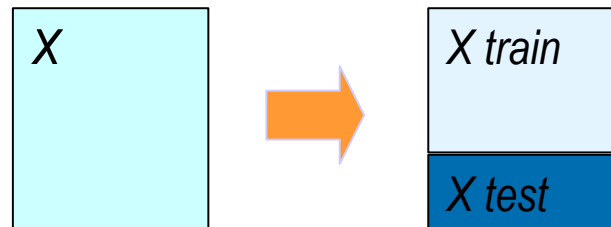
Classifier Evaluation

- The performance of your classifier needs to be evaluated on your test data:
 - an independent "validation cohort"
 - or retain a set of samples (1/3) that has similar distribution of classes of your total data



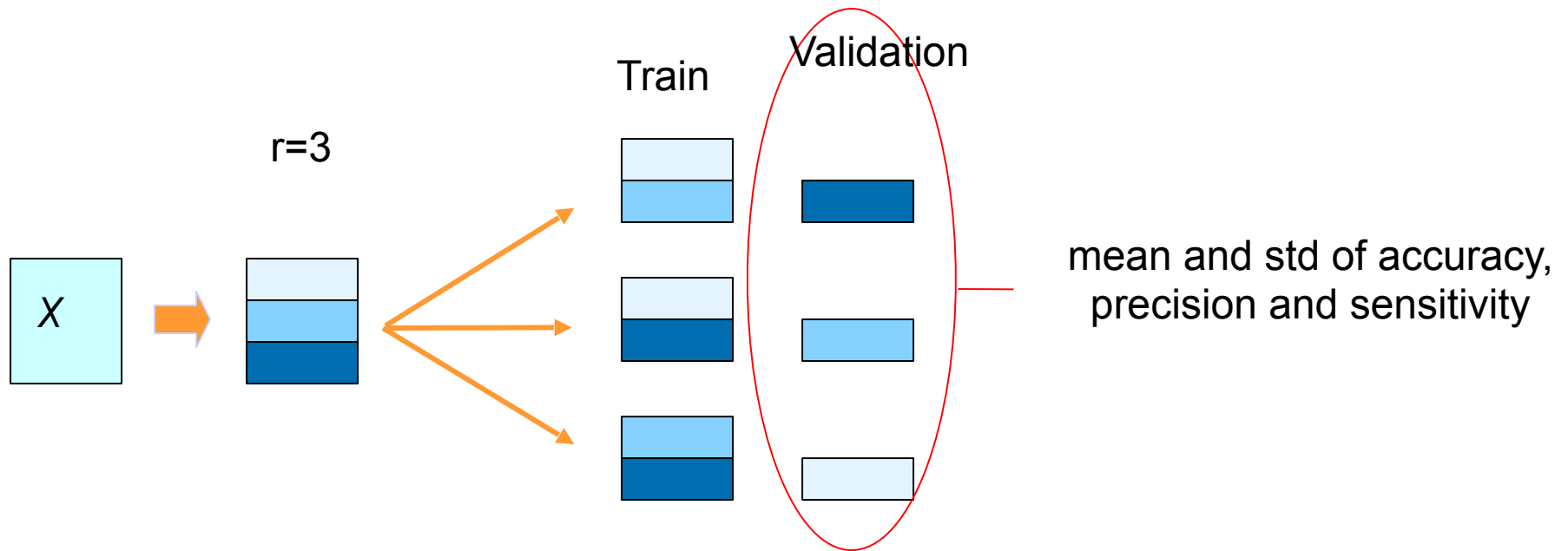
Classifier Evaluation

- The performance of your classifier needs to be evaluated on your test data:
 - an independent "validation cohort"
 - or retain a set of samples (1/3) that has similar distribution of classes of your total data



- Never use test data to improve classification (choose a better classifier or marker gene)
 - For this you need to establish validation data (or cross validation)

Cross-validation



Elastic Net

Is based on a linear function:

$$f(x, A) = a_0 + a_1x_1 + \dots + a_Lx_L$$

$$f(x, A) > 0 \Rightarrow \text{classe A}$$

$$f(x, A) \leq 0 \Rightarrow \text{classe B}$$

- Find coefficients A , *while most of them have 0*.
 - A shrinkage factor (λ) controls the number of genes selected.
 - Shrinkage factor can be automatically identified with cross-validation.

Hands on!

Exercise (after the handout)

You should perform clustering of tissues with liver cancer. Tip: use code similar to the one seen in gene expression data (day 3). Since, we are interested in grouping patients, you can transpose the matrix with the function `t`.

1. Can you see nice clusters in the dendrogram?
2. What about genes associated to each group? Are they associated to some particular biological function? Use differential expression analysis and GO enrichment analysis to solve this task.



www.costalab.org

Institute for
Computational Genomics
01011011010
1010010010

RWTHAACHEN
UNIVERSITY

Survival Analysis

- Can be used to evaluate if characteristics of a patients indicates an increase/decrease risk of survival
- clinical: tumour type, gender
 - Molecular: expression of a gene, mutation

Common Survival Tests:

- Cox proportional hazards regression (not seen here)
 - Compares survival with a numeric variable
- Kaplan-Meier graph / Log-rank test
 - compares the survival of groups of individuals

Kaplan-Meier graph / Log-rank test

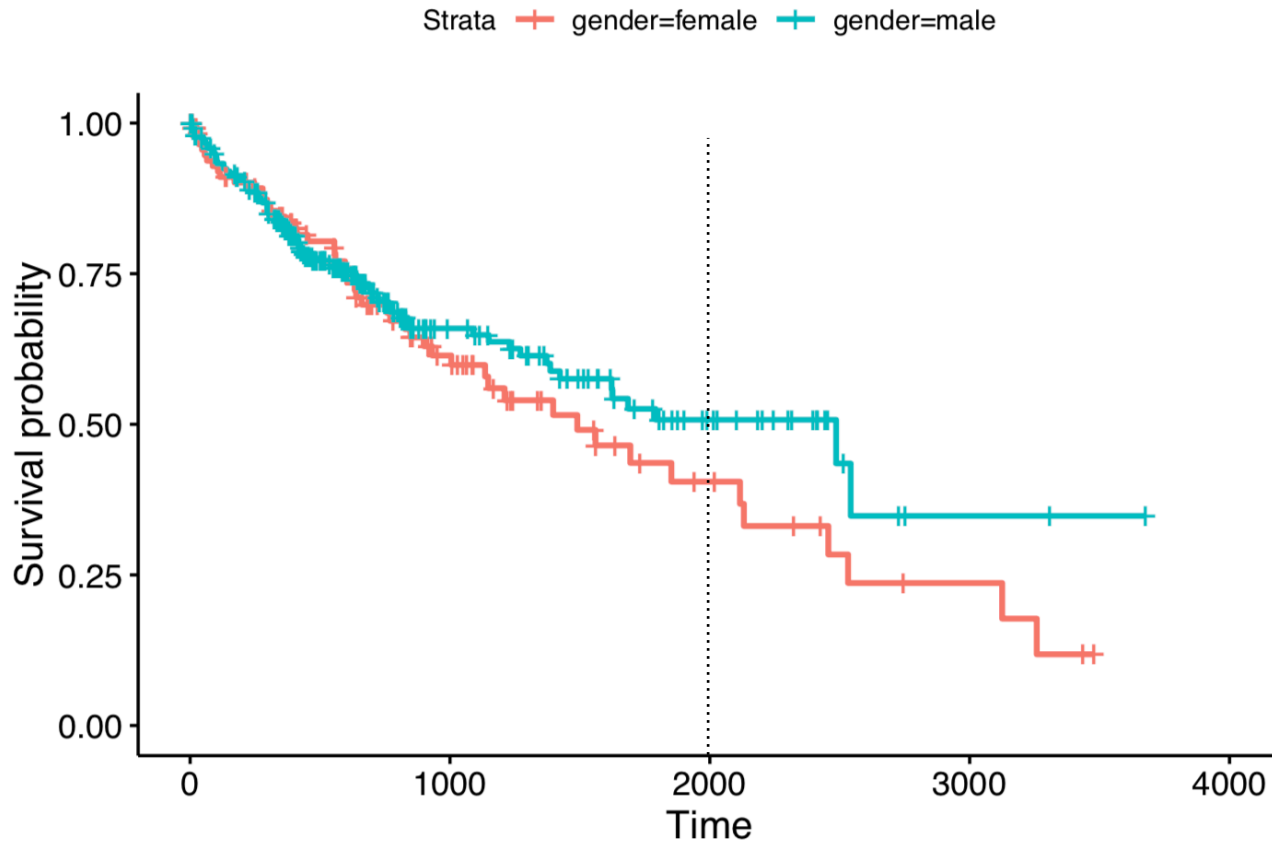
Data:

- **Event:** death / alive
- **Time:** period between first and last observation.
- **Characteristics:** sex, tumor grade

<i>Patient</i>	<i>Status</i>	<i>Time</i>	<i>Sex</i>
<i>1</i>	<i>Dead</i>	<i>343</i>	<i>Male</i>
<i>2</i>	<i>Alive</i>	<i>20</i>	<i>Male</i>
<i>3</i>	<i>Alive</i>	<i>300</i>	<i>Female</i>
<i>4</i>	<i>Dead</i>	<i>200</i>	<i>Male</i>

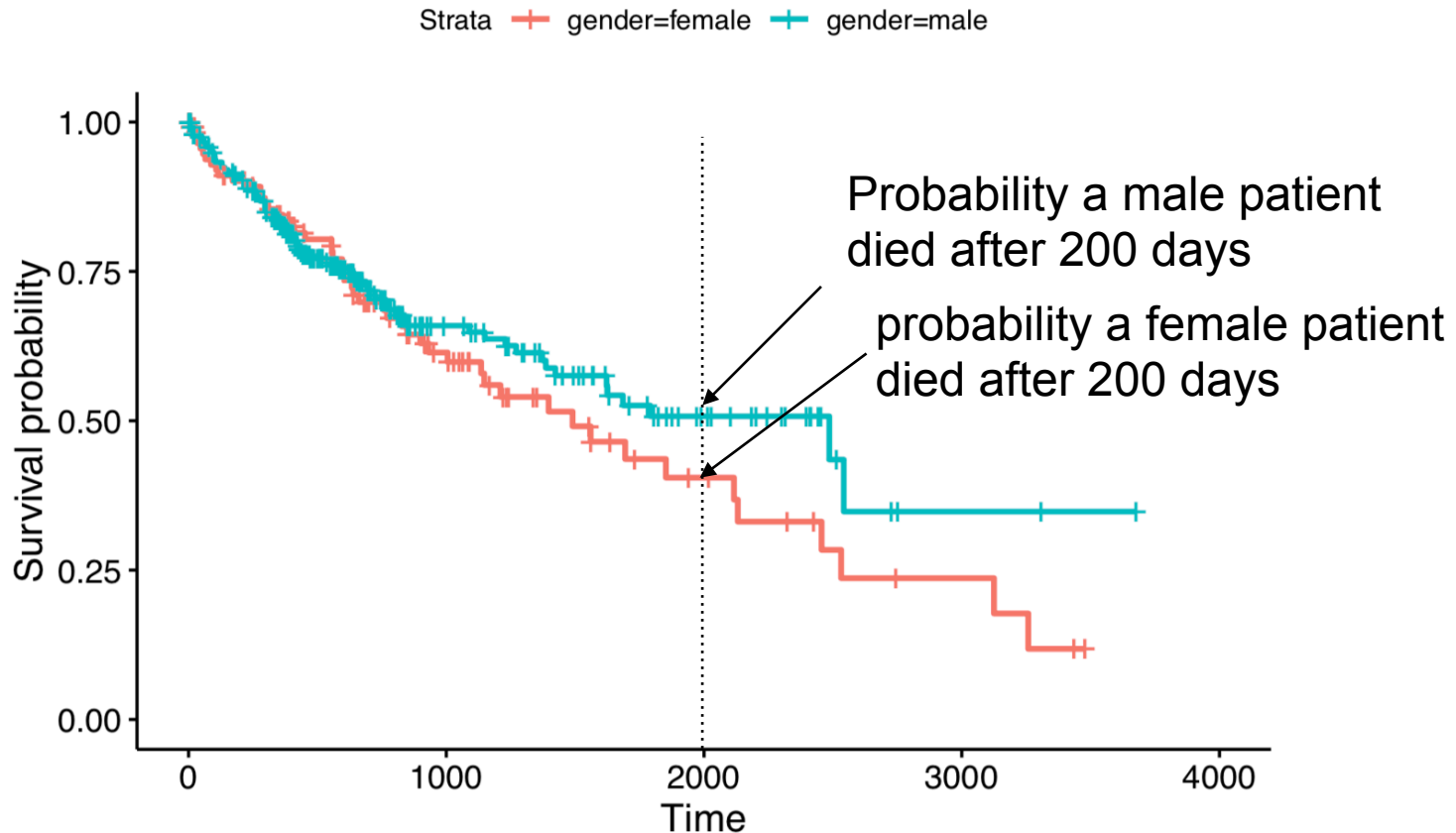
Kaplan-Meier plot

Survival of LHC patients - male vs. Female



Kaplan-Meier plot

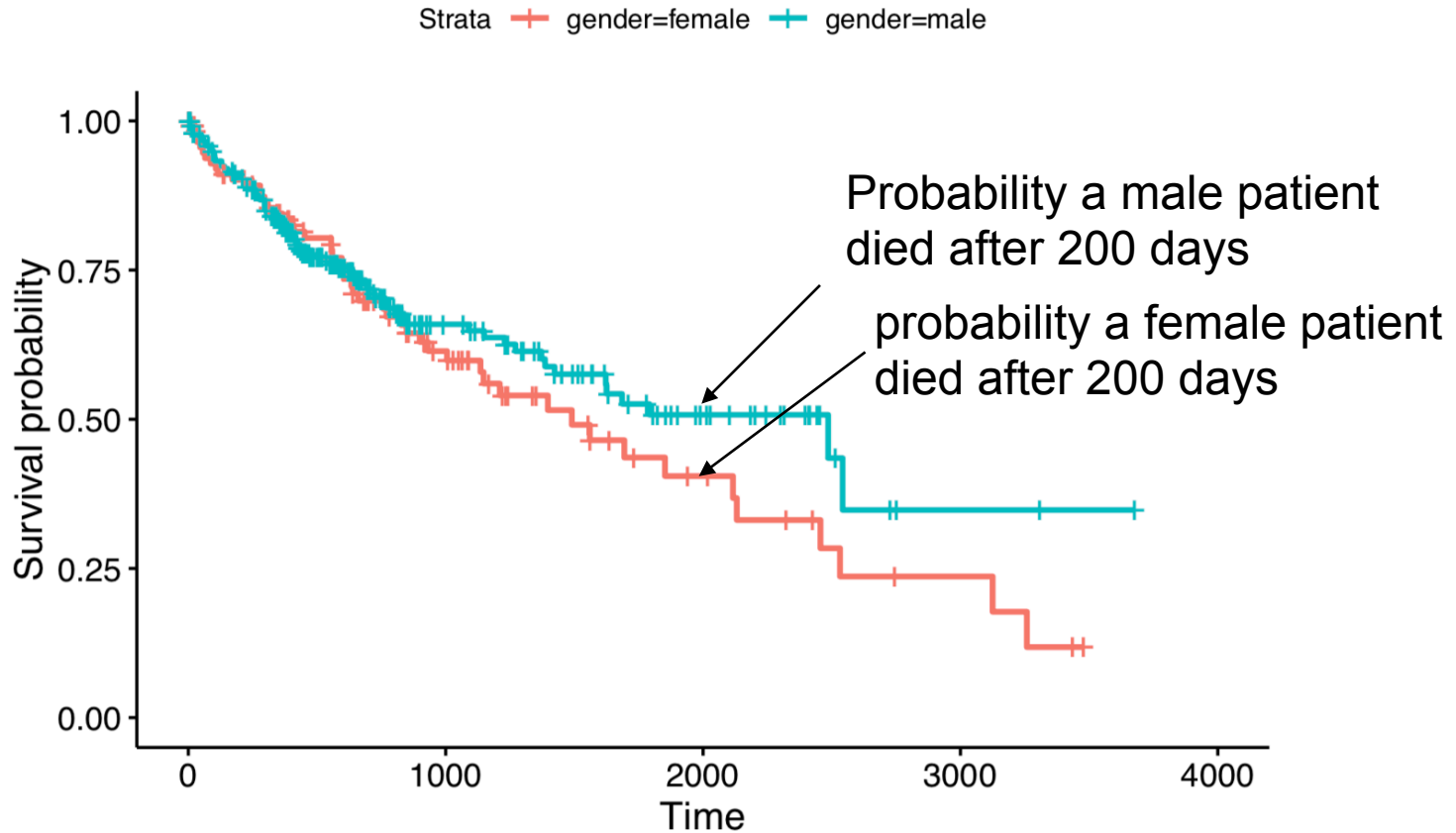
Survival of LHC patients - male vs. Female



$$\text{Probability (X days)} = \frac{\# \text{ cases alive after X days}}{\# \text{ cases measured after X days}}$$

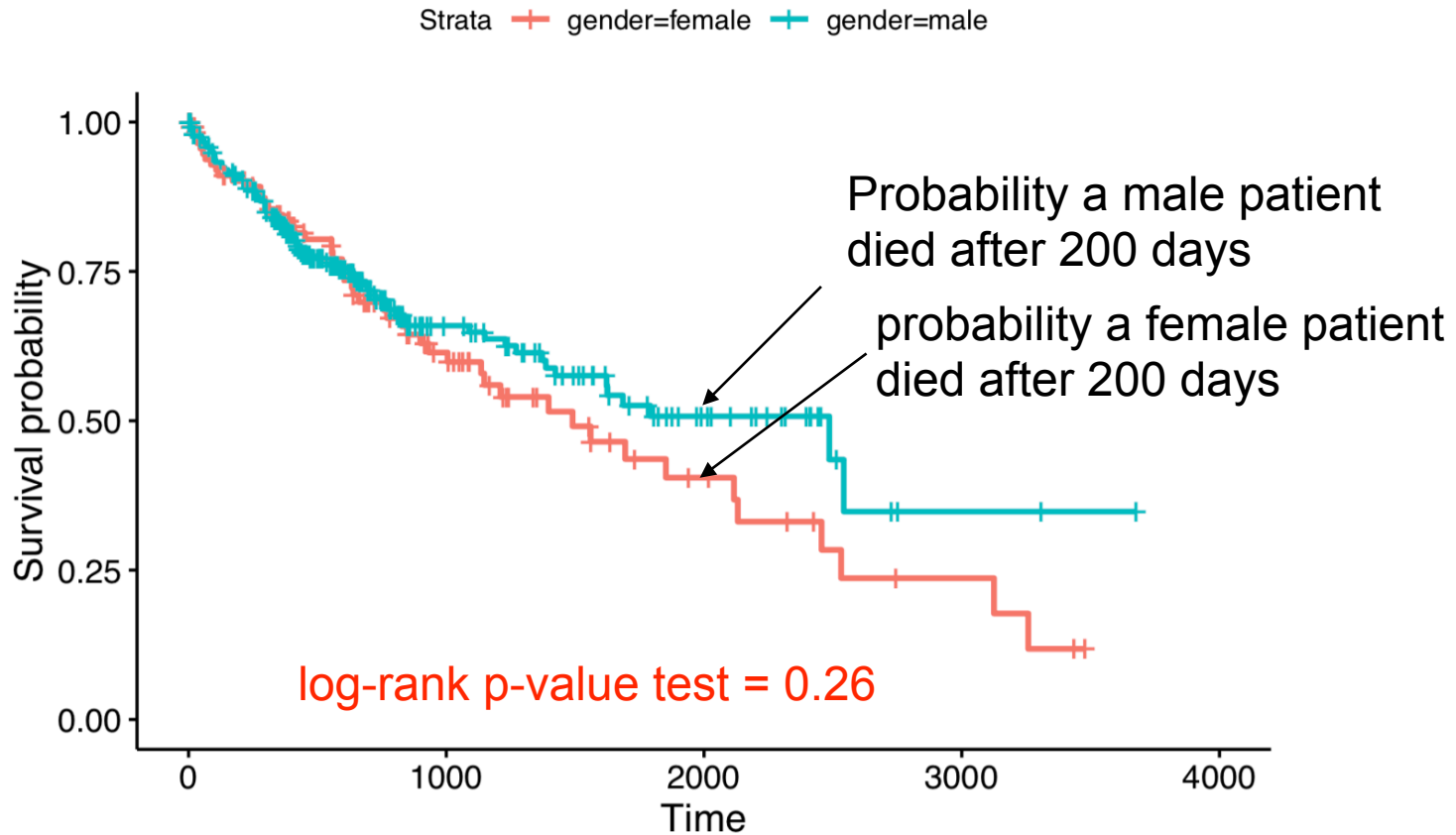
Log-rank test

Is the survival difference significant?



Log-rank test

Is the survival difference significant?



Kaplan-Meier plot

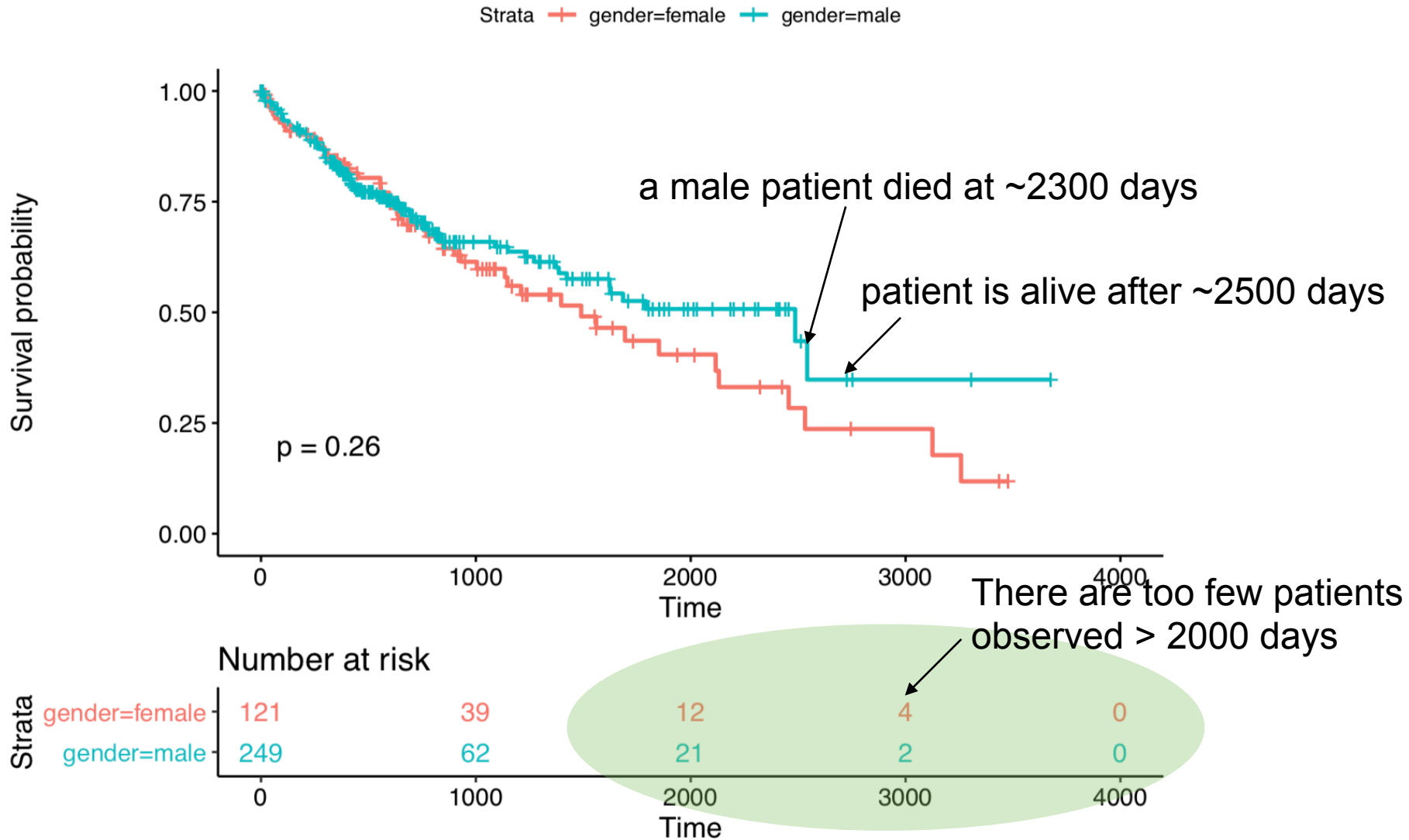


Number at risk

Strata	0	1000	2000	3000	4000
gender=female	121	39	12	4	0
gender=male	249	62	21	2	0

Time

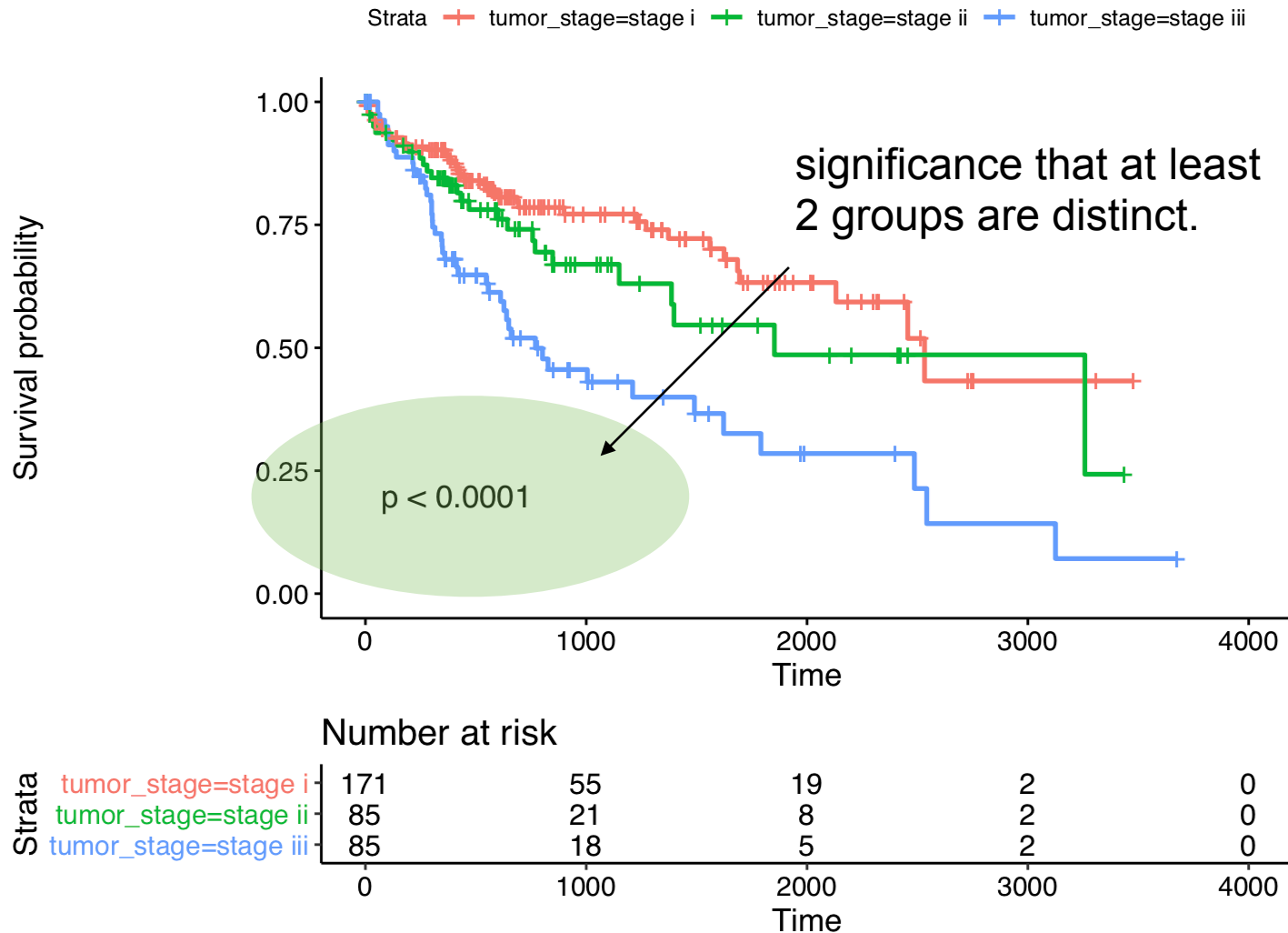
Kaplan-Meier plot



Kaplan-Meier / Log-Rank Test

KM and LRT can compare several groups at a time.

Survival vs Tumour stage at diagnosis



Survival Analysis and Biological Markers

How to perform survival analysis on biological markers?

1. Given their continuous nature of gene expression, Cox hazards test is recommended.
2. An alternative is to group patients by expression of a gene (low/high expression) and use Kaplan-Meier plots (seen in practical).

Important: if you test several markers you need to correct for multiple testing!!!

Hands on!

Exercise (after the handout)

You should perform clustering of tissues with liver cancer. Tip: use code similar to the one seen in gene expression data (day 3). Since, we are interested in grouping patients, you can transpose the matrix with the function `t`.

1. Can you see nice clusters in the dendrogram?
2. What about genes associated to each group? Are they associated to some particular biological function? Use differential expression analysis and GO enrichment analysis to solve this task.
3. **Check if group of patients are associated to survival, tumour grade or any other clinical variable? You can use the `table` function for some of these analysis.**