Bioinformatics Lab

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Objectives

- Hands on introduction to bioinformatics programming
- Review basic biological/computational aspects
 - 1. basics of molecular biology
 - 2. basics of sequencing
 - 3. basics bioinformatics problems
 - short sequences read alignment
 - gene expression quantification
 - single cell approaches
 - computational epigenetic



Objectives

- Introduction to Bioinformatics Frameworks/Tools
 - 1. biological sequence data formats/handling
 - Biopython, Pysam, R/bioconductor
 - 2. bioinformatics tools
 - BWA (aligner), Seurat, Cell Ranger, ...



Grading/Online material

Evaluation:

- 20% prototypes
- 60% final project
- 20% presentation

Extra-work for media informatics:

research report

References/Courses Online

http://costalab.org/teaching/bioinformatics-software-lab-2023/



Introduction to Molecular Biology



Understanding Live in a Molecular Level

How is genetic information inherited?

How the genetic information influence cellular processes?

How genes work together to promote particular molecular functions?



Genetic Information - DNA

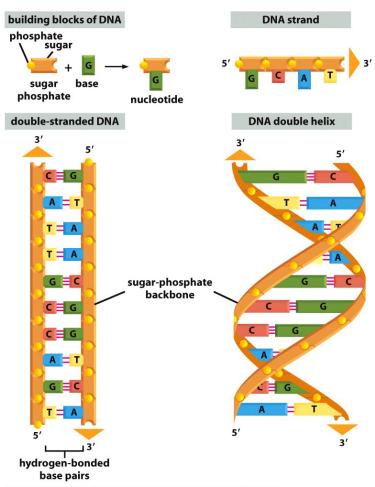
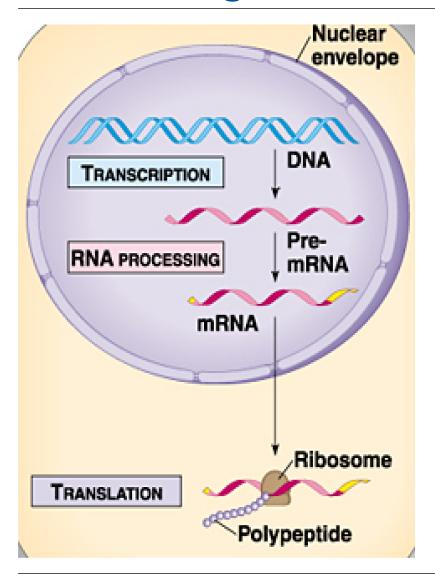


Figure 4-3 Molecular Biology of the Cell 5/e (© Garland Science 2008)

DNA (Deoxyribonucleic)

- chain of nucleic acids
- 4 bases: A;C;G;T
- forms DNA duplexes with paring A = T e C = G

Central Dogma - Transcription



Transcription

- DNA to RNA RNA (ribonucleic acid)
 - single stranded
 - 4 bases: A;C;G;U
 - unstable
 - transport of information from nucleus to cytoplasm



Central Dogma - Transcription

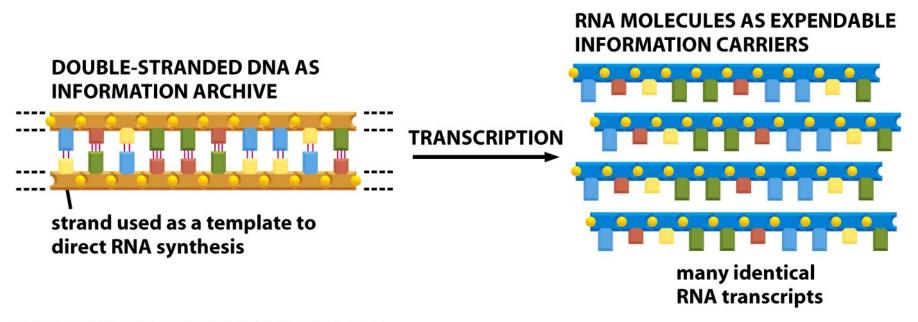
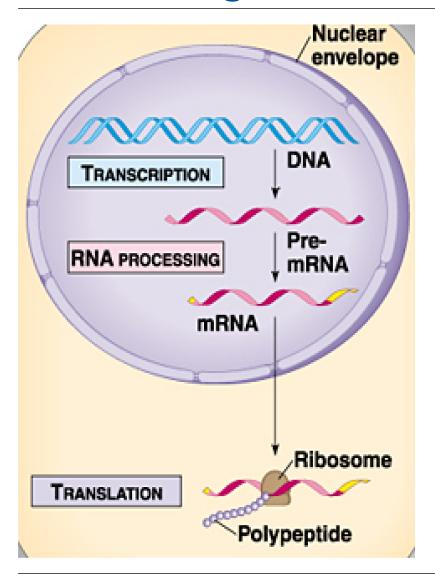


Figure 1-5 Molecular Biology of the Cell 5/e (© Garland Science 2008)

Transcription - copy of DNA information to RNA (T to U)



Central Dogma - Translation



Translation

- RNA to Protein
- performed by the ribosome
- follows the genetic code

Proteins

- single stranded chain
- 20 amino acids
- assumes 3D structure
- main functional entities in the cell



Genetic Code - Translation

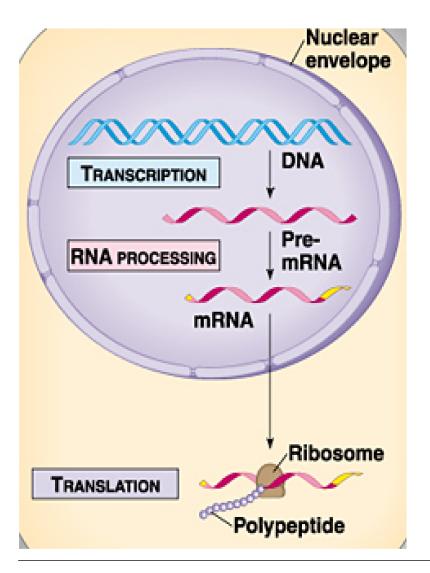
GCC GCG	AGA AGG CGA CGC CGG CGU								AUC		AAA	AUG		CCC	AGC AGU UCA UCC UCG UCU	ACC ACG	UGG		GUA GUC GUG GUU	UAA UAG UGA
Ala	Arg	Asp	Asn	Cys	Glu	Gln	Gly	His	lle	Leu	Lys	Met	Phe	Pro	Ser	Thr	Trp	Tyr	Val	stop
Α	R	D	N	C	E	Q	G	Н	1	L	K	M	F	Р	S	Т	W	Υ	V	

Figure 6-50 Molecular Biology of the Cell 5/e (© Garland Science 2008)

triples of RNA bases encodes a amino acid



Central Dogma



- Dogma: information flux
 DNA -> mRNA -> Proteins
- Gene: DNA segment coding a protein.
- Transcript: RNA segment associated to a gene.
- Genes is associated to one proteins and one function*

* Genes might be associated to many proteins



Control of Gene Expression

How is the expression of genes controlled?

enhancer
(binding site for activator protein)

BINDING OF transcription
GENERAL TRANSCRIPTION
FACTORS, RNA POLYMERASE,
MEDIATOR, CHROMATIN REMODELING
COMPLEXES, AND HISTONE ACETYLASES

Certain proteins
(transcription factors)
bind to DNA and
initiate transcription

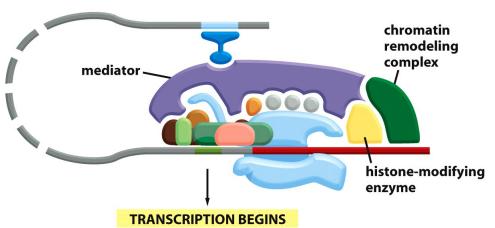
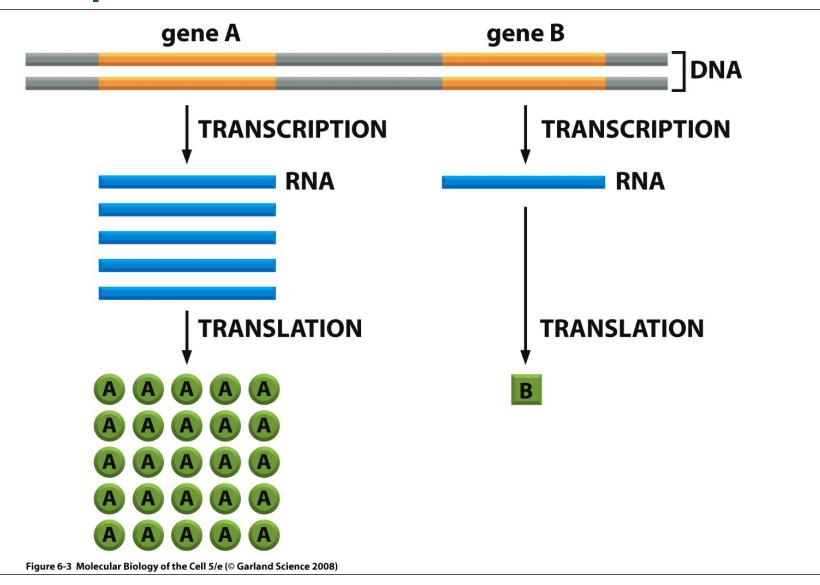


Figure 6-19 Molecular Biology of the Cell 5/e (© Garland Science 2008)



Gene Expression



Cellular Complexity

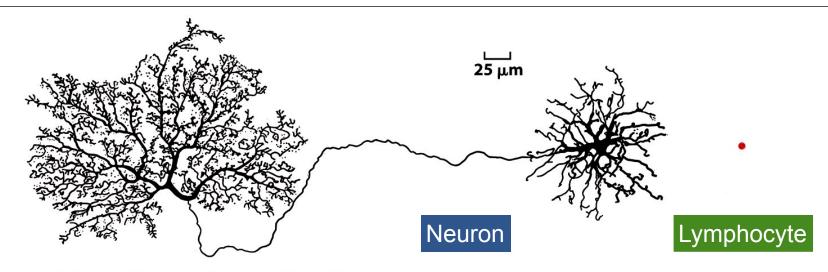


Figure 7-1 Molecular Biology of the Cell 5/e (© Garland Science 2008)

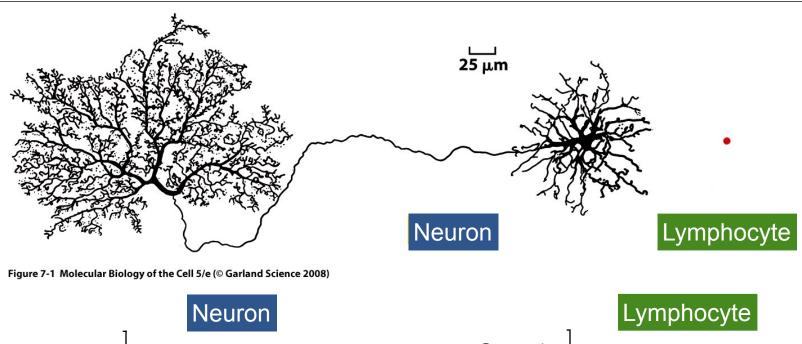
Two cells of a organism have exactly* the same DNA

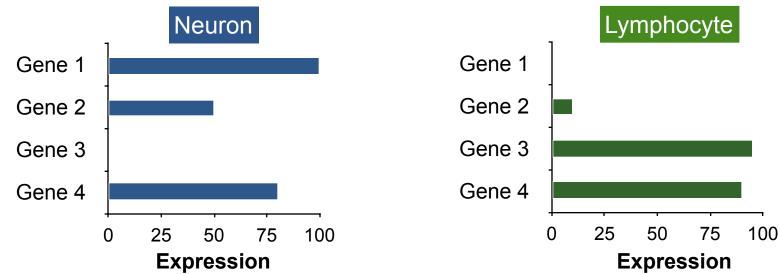
How does this differences arise? How is cell fate remembered?

* with exception of somatic mutations and rearrangements of immunological loci



Cellular Complexity & Gene Expression







Sequencing

Sequencing

Read the bases of a particular DNA/RNA sequence

Applications:

- sequence DNA of known and unknown organism
- detect variants on patients
- sequence the RNA of a cell
- detect location of proteins interacting with DNA or open chromatin

Problem:

- only short DNA sequences (<1.000 bs) can be read

Solution:

break DNA in several small pieces and use bioinformatics



Next Generation Sequencing

- NGS take advantage of parallelization
 - reads millions/billions of reads for a time
 - short reads (50-300 bps)
 - moderate error rates (0.1%)
- commercial products:
 - **454**
 - SOLID
 - Solexa (Illumina)



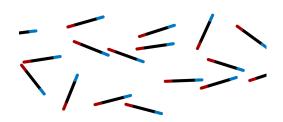


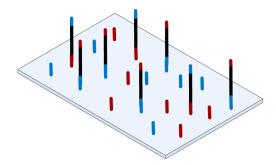
Illumina Flow Cell - NGS Sequencing

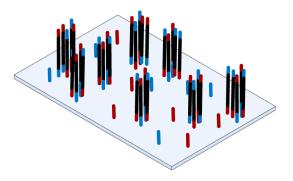
1- fragment sample DNA, insert adapters, attach to flow cell

2- use (bridge) PCR to copy fragments (close to origin)

3- clusters of single stranded DNA (200m clusters with 2k DNA strands





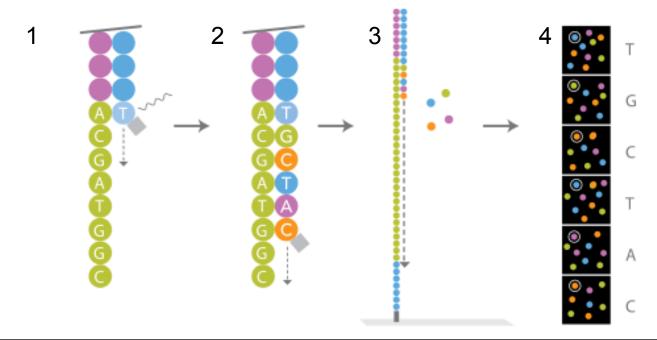


See video http://www.wellcome.ac.uk/Education-resources/Education-and-learning/Resources/Animation/WTX056051.htm

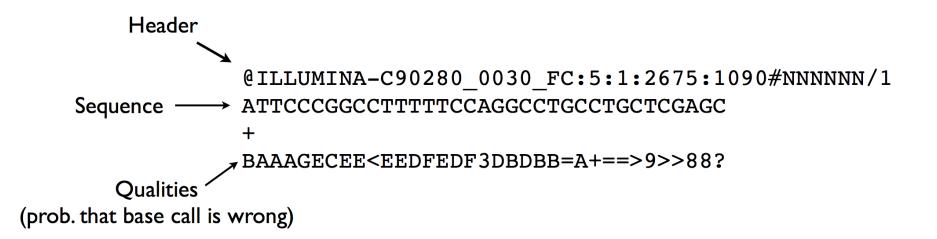


Illumina Flow Cell - NGS Sequencing

- Iterative evaluation process:
 - 1. add RT-bases, polymerases integrate them
 - 2. wash away all not integrated elements
 - 3. take picture of flow cell to determine current base by dye
 - 4. derive reads from pictures



Sequencing Results



One character encodes a number using ascii table (0-255)

This number (Q) can be converted to P

Phred-scale

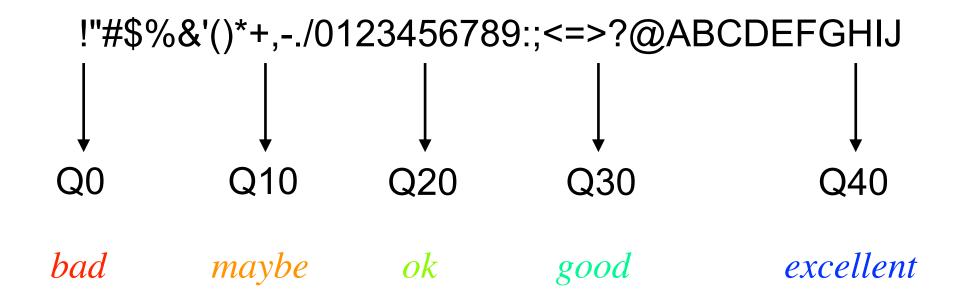
Q = -10 * log 10 P

 $P = 10^{(-Q/10)}$



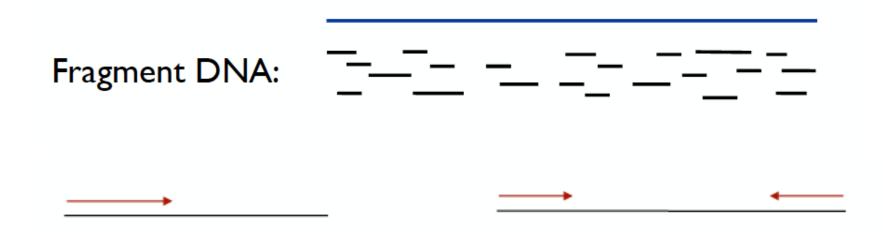
Sequencing Results / Phred scores

Uses letters/symbols to represent numbers:





Read Types



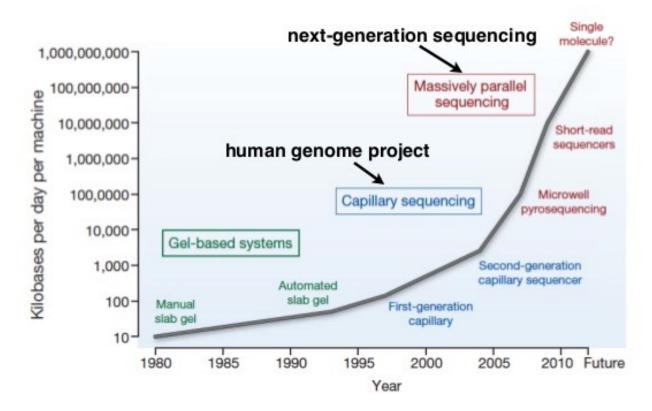
Single end

Paired end Ins: 200-800 bp



Next Generation Sequencing

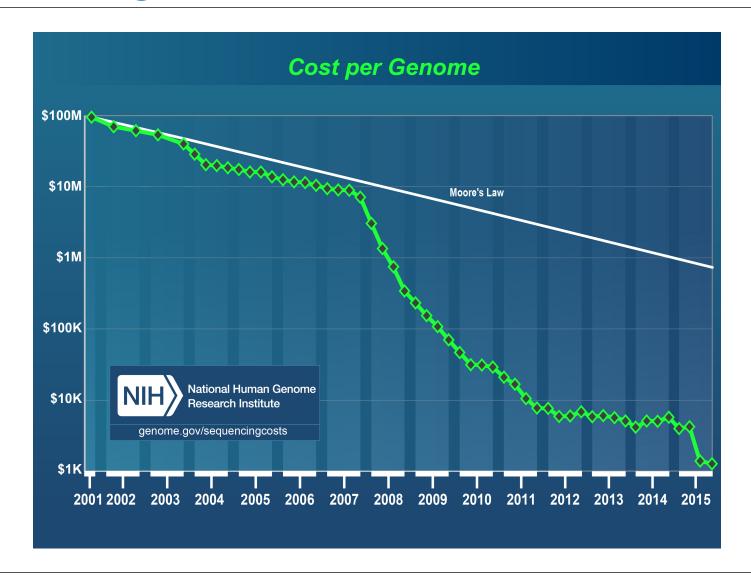
Improvements in the rate of DNA sequencing over the past 30 years



Stratton, M. R., Campbell, P. J. & Futreal, P. A. The cancer genome. Nature 458, 719-724 (2009).



Sequencing Costs





Sequence Alignment



Sequence Alignment

NGS

- reads from DNA fragments
- position in genome is unknown
- solution: alignment

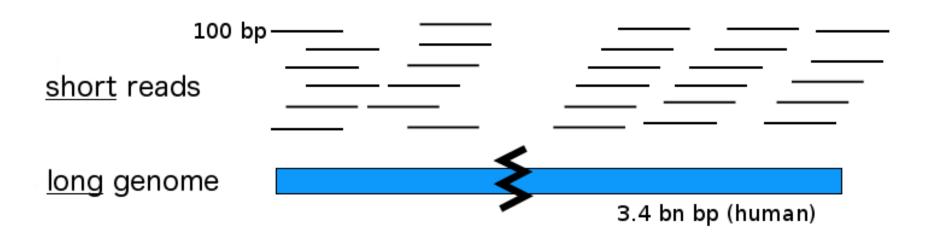
DNA Sequencing

- de-novo assembly
 - construct unknown reference sequence from scratch
- resequencing / mapping
 - reference sequence given (applies to human- and mousestudies)
 - build sequence that is similar but not necessarily identical to reference sequence



Alignment Problem

- a large reference sequence is given (genome)
 - up to billions of base pairs
- millions of short reads (<200bps)
- find most probable position of the read in the genome (by inexact string matching)





Pitfals

- (Unknown) divergent of sample and reference genome
- Repeats in the genome (larger than read size)
- Recombinations
- Poor genome reference quality
- Sequencing/read errors



Alignment/Mapping is a typical inexact string match problem

Algorithmic Solutions: ?



Alignment/Mapping is a typical inexact string match problem

Algorithmic Solutions:

 Smith & Waterman - dynamic programming (quadratic time/memory)



Alignment/Mapping is a typical inexact string match problem

Algorithmic Solutions:

- Smith & Waterman dynamic programming (quadratic time/memory)
- Blast k-mer search for seeding followed by dynamic programming
 - large memory requirement
 - local alignment



Short read alignment is a special problem

- reference sequence is large and fixed
- query sequence (reads) are short and many

Solution: ?



Short read alignment is a special problem

- reference sequence is large and fixed
- query sequence (reads) are short and many Solution: ?
- 1. Use a data structure to represent reference
 - k-mer hash table (>40GB for k=8)
 - suffix trees (> 4GB)

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- 2. Find candidate (k-mer) hits on genome (>100)



Short read alignment is a special problem

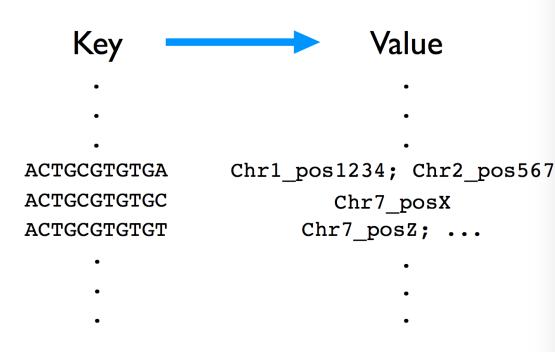
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- query sequence (reads) are short and many
 Solution: ?
- 1. Use a data structure to represent reference
 - k-mer hash table (>40GB for k=8)
 - suffix trees (> 4GB)
- 2. Find candidate (k-mer) hits on genome (>100)
- 3. Improve alignment with Smith-Waterman Methods work on linear time (query sequence)



Hash based algorithm

Lookups in hashes are fast!

- I. Index the reference using *k*-mers.
- 2. Search reads vs. hash k-mers
- 3. Perform alignment of entire read around seed
- 4. Report best alignment

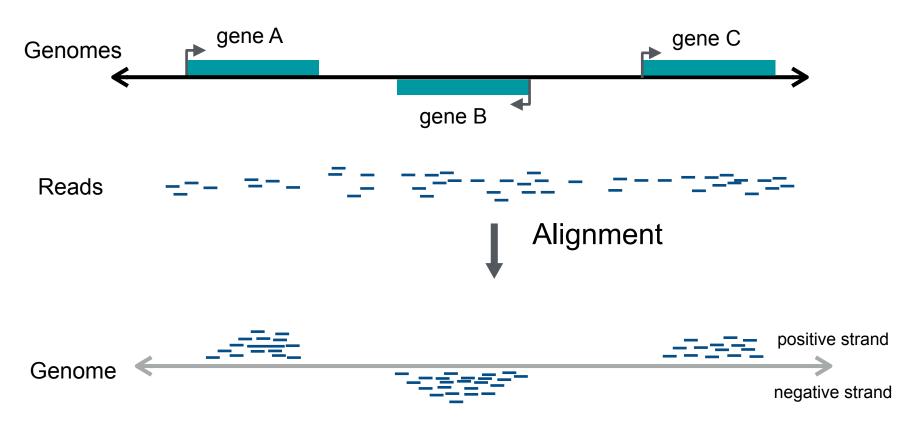


Also known as Seed and extend



RNA sequencing / Alignment Results

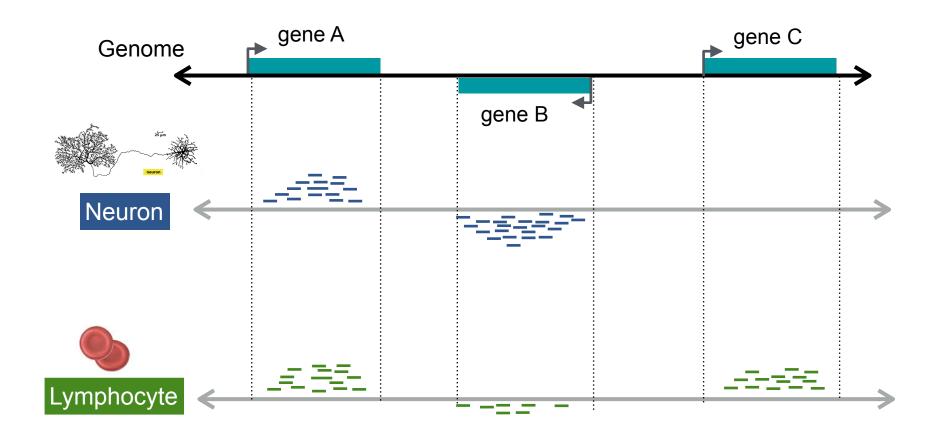
- Position and strand of reads aligned to the genome





Gene Quantification

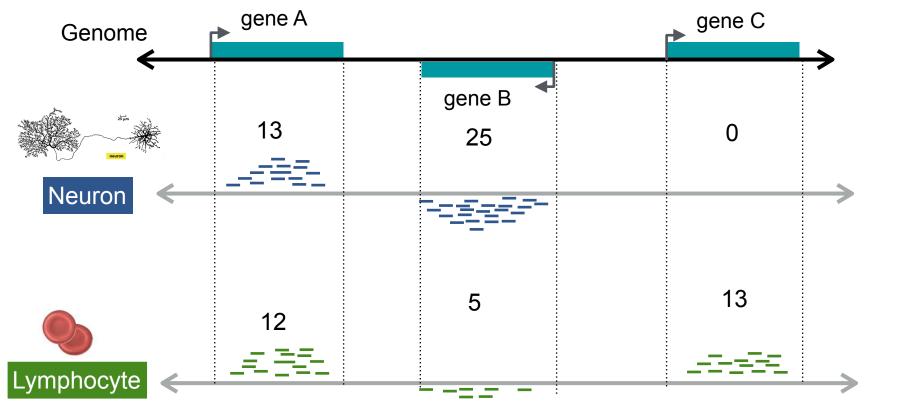
- Perform sequencing for each cell (neuron, lymphocyte)
- Align reads to genome





Gene Quantification

- Perform sequencing for each cell (neuron, lymphocyte)
- Align reads to genome
- Count number of reads inside genes (using known genes annotation)





Quantification - Normalization

Correct for:

- Genes having distinct size
- Sequencing efficiency differs between cell (usually same RNA quantity provided for sequencing)

	Cell A	Cell B	
GeneA (1kb)	20	15	30
GeneB (2kb)	100	300	10
GeneC (1.5kb)	10	20	100
Gene D (3kb)	300	200	100
Total Library	430	535	240

Reads per kilobase million (RPKM) = #reads * gene size * total library

1.000 1.000.000



Resume

- Review basic biological/computational aspects
 - 1. basics of molecular biology
 - 2. basics of sequencing
 - 3. basics bioinformatics problems
 - short sequences read alignment
 - gene expression quantification
 - single cell sequencing (next)
 - computational epigenetic (next weeks)

